

Internal Coordinate Mechanics and ICM

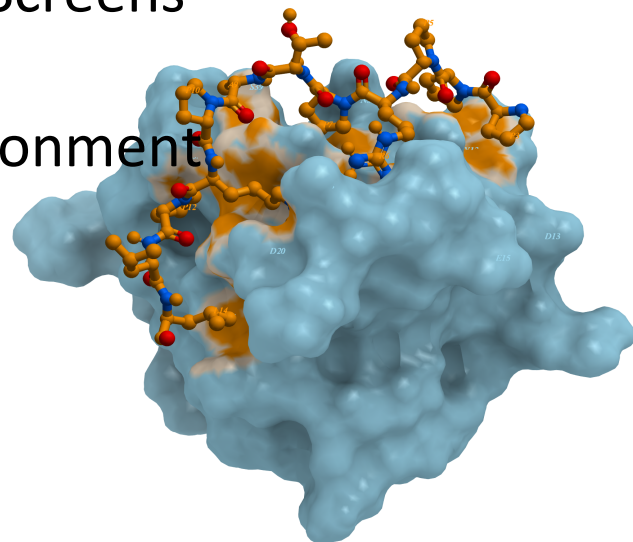
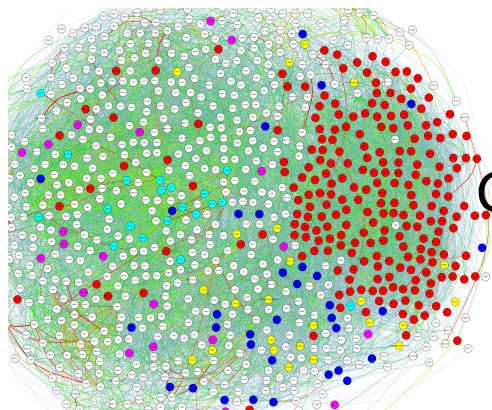
Modeling

Docking, Ligand and Target Screens

Cheminformatics

Graphical and Scripting Environment

Big Structural Data



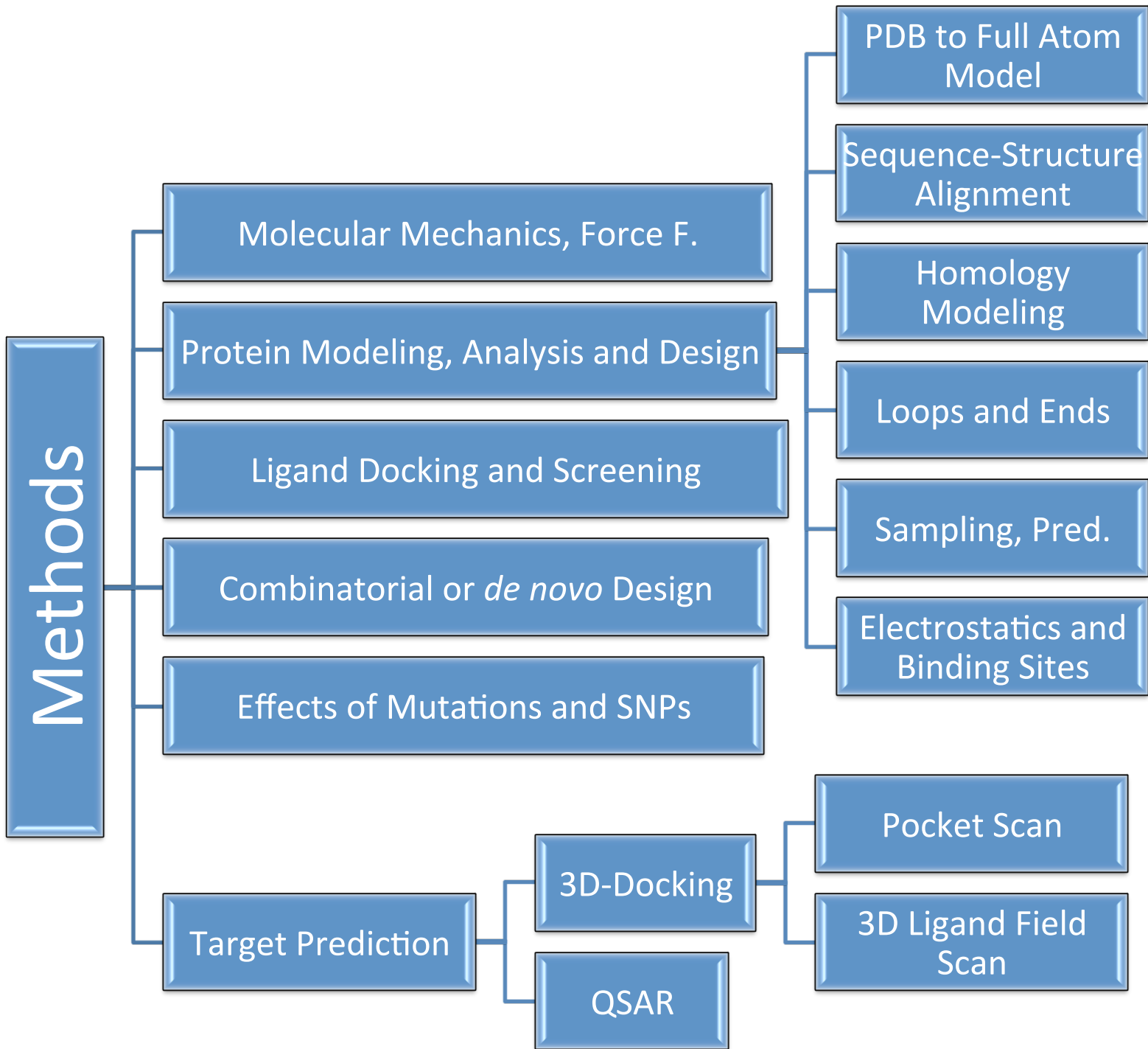
Ruben Abagyan

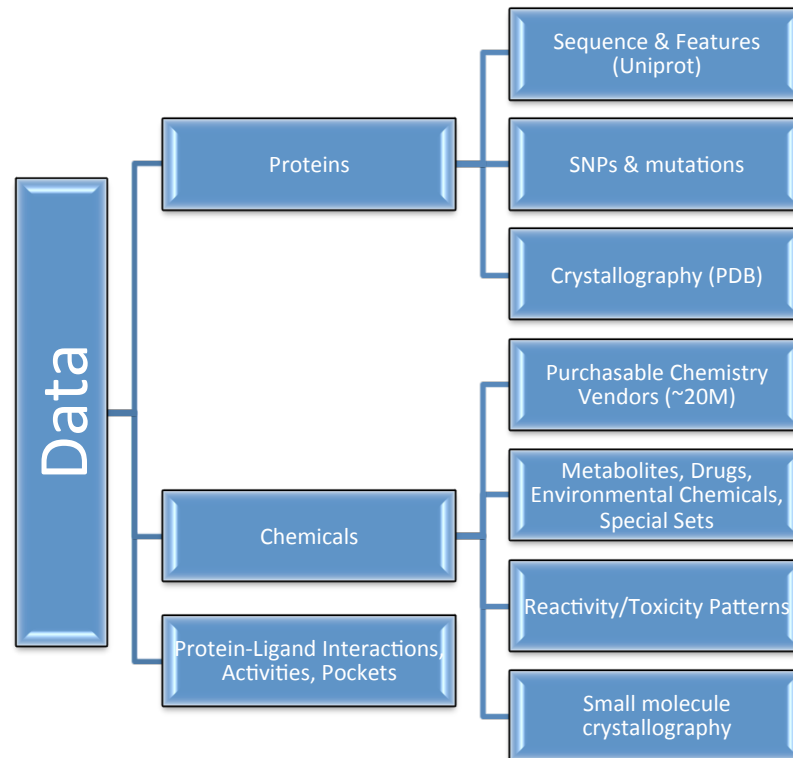
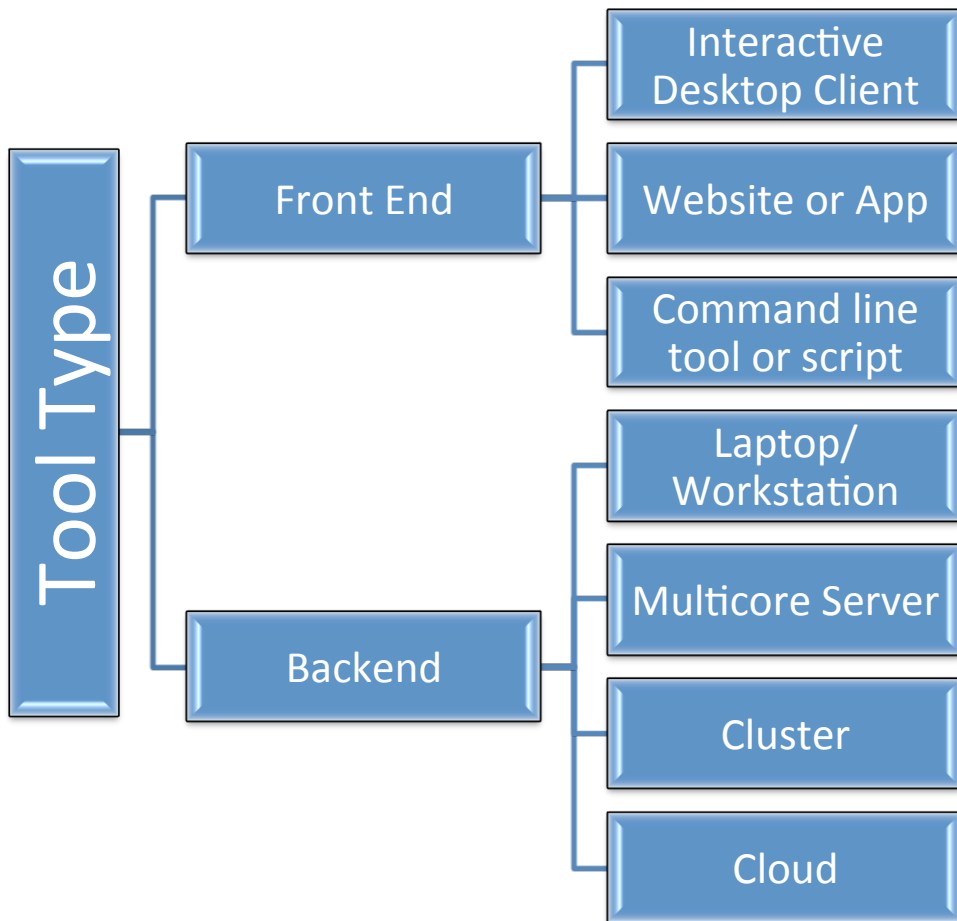
University of California, San Diego

Skaggs School of Pharmacy & Pharm. Sciences

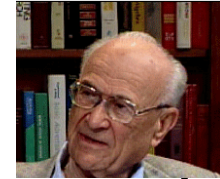
Molsoft, LLC

ruben@ucsd.edu





Internal Coordinate Mechanics for modeling&docking biological complexes



Harold Scheraga

Energy, FF

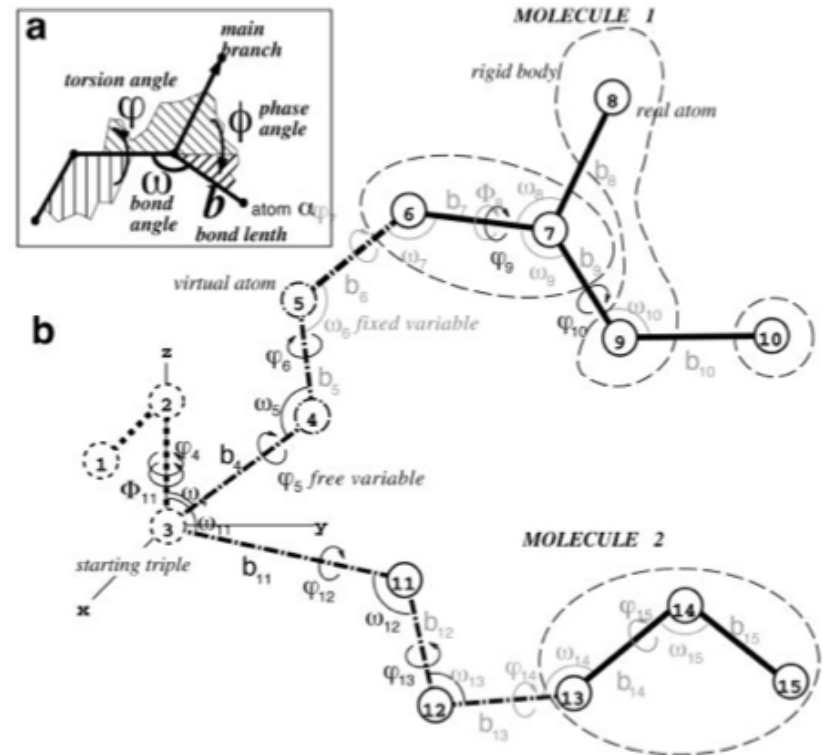
\mathbf{X} , $E(x, y, z)$

\mathbf{Q} $E(\varphi)$
 $E(b, \alpha, \varphi, \phi)$

Dynamics

Newton

$$F = ma$$



Stochastic sampling



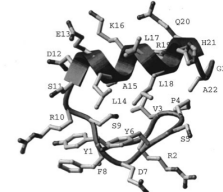
Inefficient,
 Sample in IC

Square-Root Sampling

Ab initio folding of peptides with the Optimal Bias Monte Carlo Minimization Procedure

Abagyan, Totrov *J. Comp. Physics*, 1999

Optimal Sampling: *discrete states*



The lowest energy conformation, $E = -385.4$ kcal/mol

THEOREM. *The optimal random guessing strategy minimizing the average number of guesses is to guess with relative frequencies f_1 and f_2 so that*

$$\frac{f_1}{f_2} = \sqrt{\frac{S_1}{S_2}}.$$

Proof. Let us calculate the average number of guesses N_{guesses} until the correct answer is given, provided that previous guesses are forgotten and thus each guess is independent. If in our random guessing strategy the probability of a correct guess in a single trial is f , and the successful result can be achieved through the first correct guess ($p_1 = f$), the first incorrect guess and the second correct guess ($p_2 = (1 - f)f$), the first two incorrect guesses and the correct guess ($p_3 = (1 - f)^2 f$), etc., the average number of guesses reads

$$\begin{aligned} N_{\text{guesses}} &= p_1 + 2p_2 + 3p_3 + \dots \\ &= f + 2(1 - f)f + 3(1 - f)^2 f + \dots + n(1 - f)^{n-1} + \dots = 1/f, \end{aligned}$$

Sampling function

is

chapter in

Applications, Kluwer

$$\int_{-1}^1 f_j^{-1}(x_i^0) dx_i^0,$$

$$\langle N_{\text{guesses}} \rangle = S_1/f_1 + S_2/f_2.$$

let us set the derivative of $\langle N_{\text{guesses}} \rangle$ to zero,

$$S_1/f_1^2 - S_2/f_2^2 = 0,$$

$$S_1/f_1^2 = S_2/f_2^2,$$

$$f_1/f_2 = (S_1/S_2)^{1/2}.$$

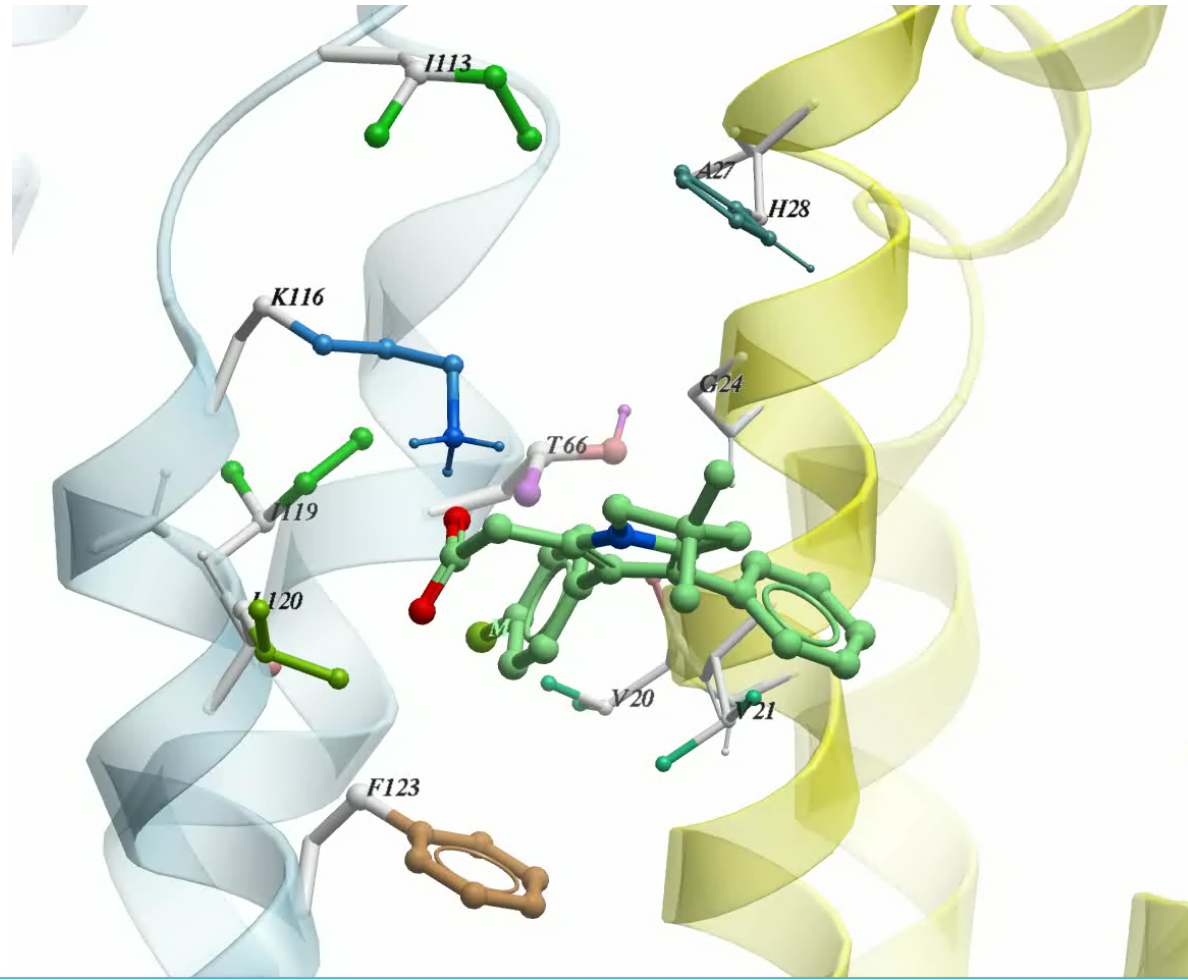
order for the above equation to hold for any arbitrary function δf_j , we have

$$f_j(\mathbf{x}) = \frac{1}{c'} \sqrt{S_j(\mathbf{x})},$$

where c' is the normalization constant equal to $\int \sqrt{S_j(\mathbf{x})} d\mathbf{x}$.

Fast Conformation Sampler and Optimizer in Internal Coordinates

- A minimal subset of internal variables
- Collective moves
- Optimal Square-root sampling
- Stochastic global optimizer with history feedback

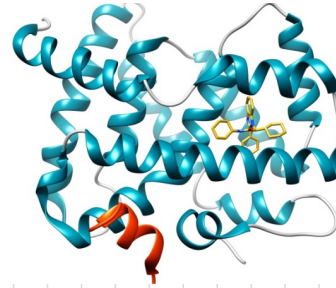


Abagyan R., Totrov M. 1994, *ICM JCC*, *BPMC JMB*, 1997 *Ligand Docking, Proteins*.

Neves et al., 2011: Top scoring pose: 91% <2Å cognate docking for 165 tasks, 71% <1Å

Bottegoni G et al. ..multiple receptor conformations for VLS. *PLoS One*, 2011, *Activity Cliffs* 2015

D3R Grand Docking Challenge: 2017

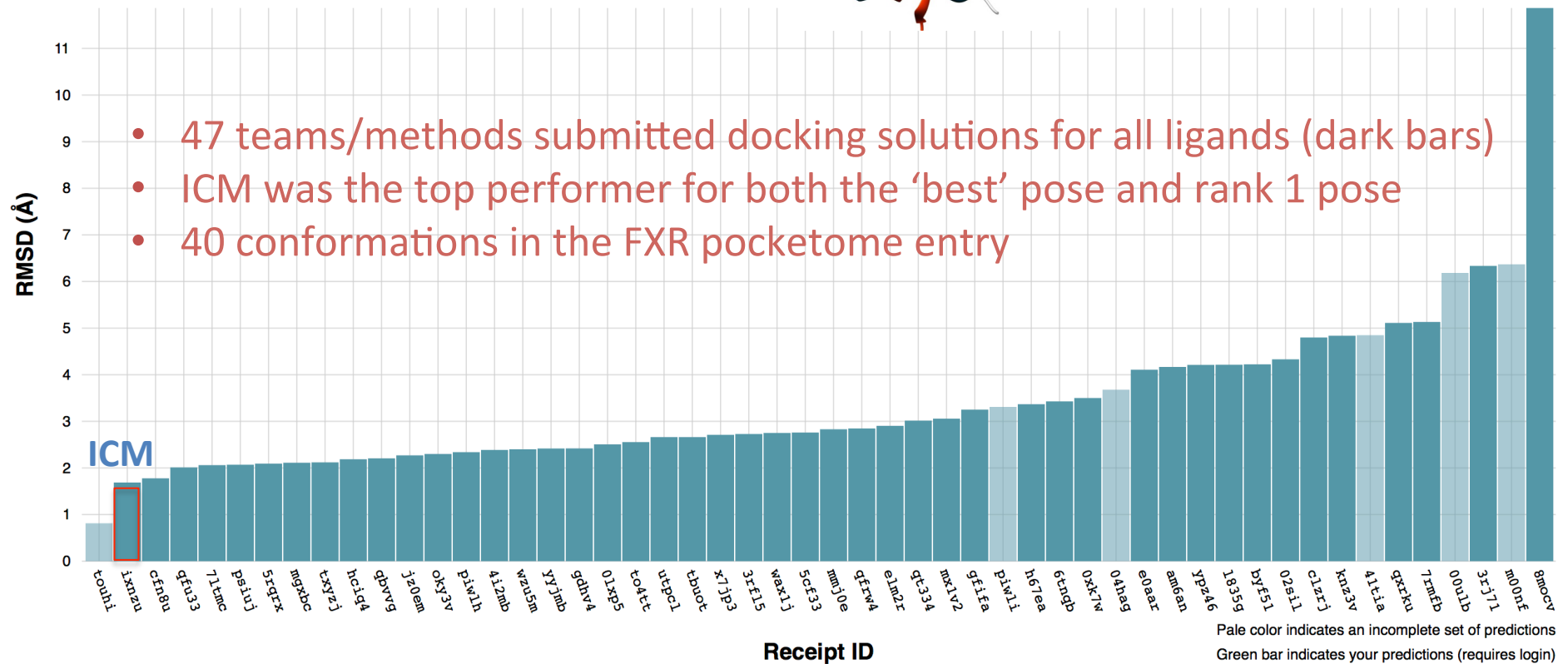


Predict the crystallographic poses of 36 ligands to FXR

Grand Challenge 2

Pose - RMSD - Compound: AVG - Best

Compound: AVG



Docking: 7 Types of Pocket Flexibility

- Explicit Side Chains in LigEdit and _dockScan
- 4D Docking / Ensemble Docking
- SCARE (SCan Alanines & Refine)
- NMA or ICM **Ligand-guided** pocket variations
- Full ICM simulations with custom defined space and restraints
- Hybrid protocols



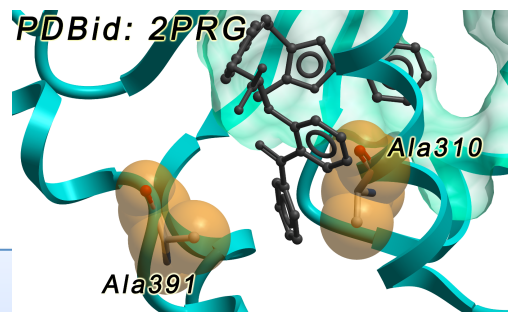
Max



Giovanni



Irina

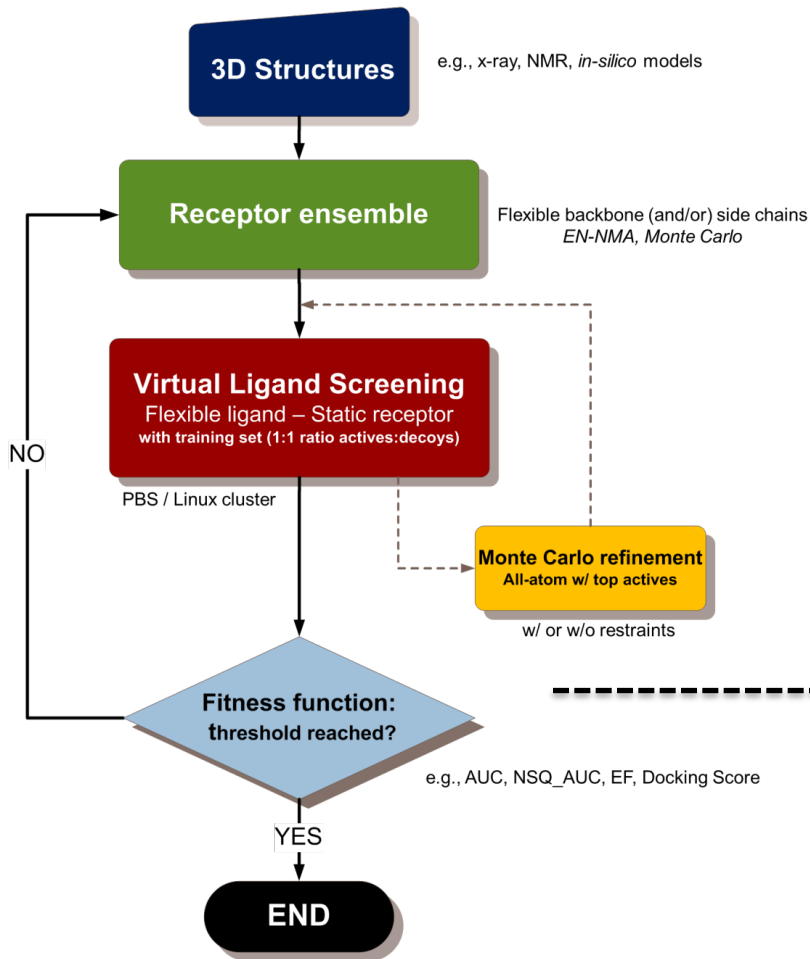


Bottegoni et al. 2008 “**SCARE..**” *JCAMD*

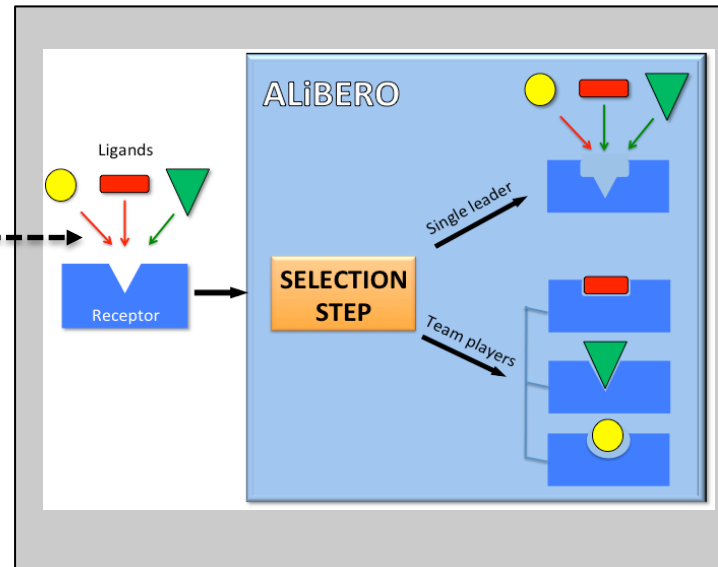
Bottegoni et al. 2009 “**4D docking: a fast and accurate account of discrete receptor flexibility ..**”, *JMC*

Husby, Bottegoni, Kufareva, Abagyan, Cavalli. **Structure-based predictions of activity cliffs.** *J Chem Inf Model.* 2015

Ligand Guided Model Building



- ALibero
- Extension of _dockScan

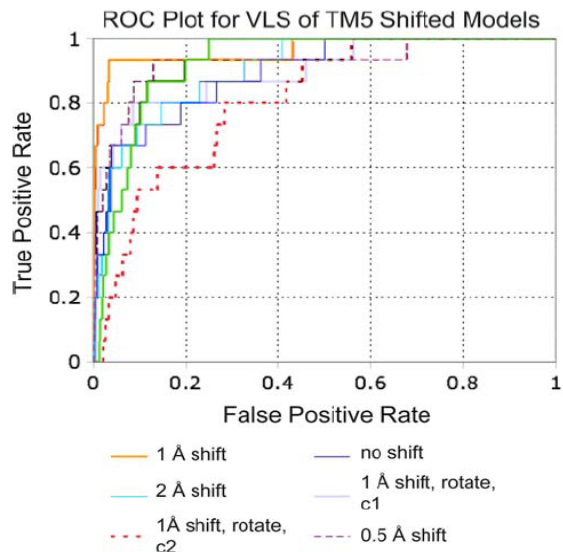


Bisson, Cheltsov et al. 2006, PNAS

Katritch et al. 2008, 2011, ++ GPCR agonist binding revealed by modeling..

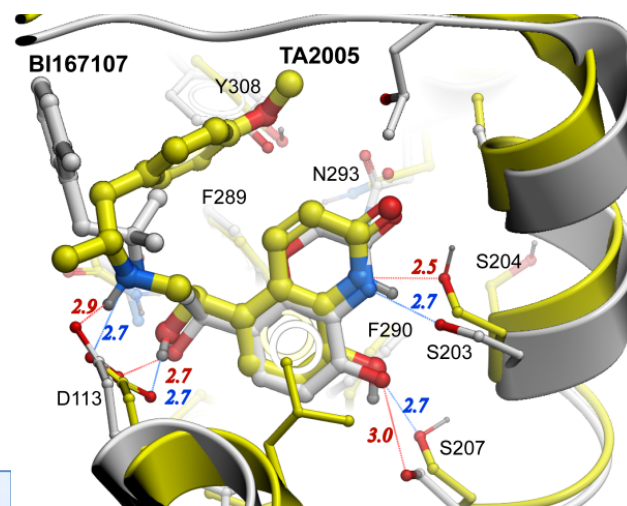
Rueda et al. **ALiBERO: Evolving a team of complementary pocket conformations rather than a single leader** (2012) *J Chem Inf Mod*

Prospective Prediction of Agonist-Bound Pocket and Agonist Binding by ICM LGM Model 2007-09, X-ray 2011



- Reynolds, Katritch, Abagyan, **Identifying conformational changes of the β_2 adrenoceptor that enable accurate prediction of ligand/receptor interactions and screening for GPCR modulators**, *JCAMD*, 2009
- Katritch, Reynolds, Cherezov, Hanson, Roth, Yeager, Abagyan. **Analysis of full and partial agonists binding to beta(2)-adrenergic receptor suggests a role of transmembrane helix V in agonist-specific conformational changes** *J Mol Recognit.* 2009 Apr 7;22(4):307-318
- Katritch V, Abagyan. **GPCR agonist binding revealed by modeling and crystallography**, *Trends PharmacolSci*, 2011 Sep 6
- Warne, et al., Schertler G, Tate C, **The structural basis for agonist and partial agonist action on a β_1 adrenergic receptor**, *Nature*, 2011.

β_2 AR agonists comparison 4 years later



100% identical contacts for ligand core
 $Rmsd_{LIG_CORE} = 0.5 \text{ \AA}$,
 $RMSD_{pocket} = 0.9 \text{ \AA}$

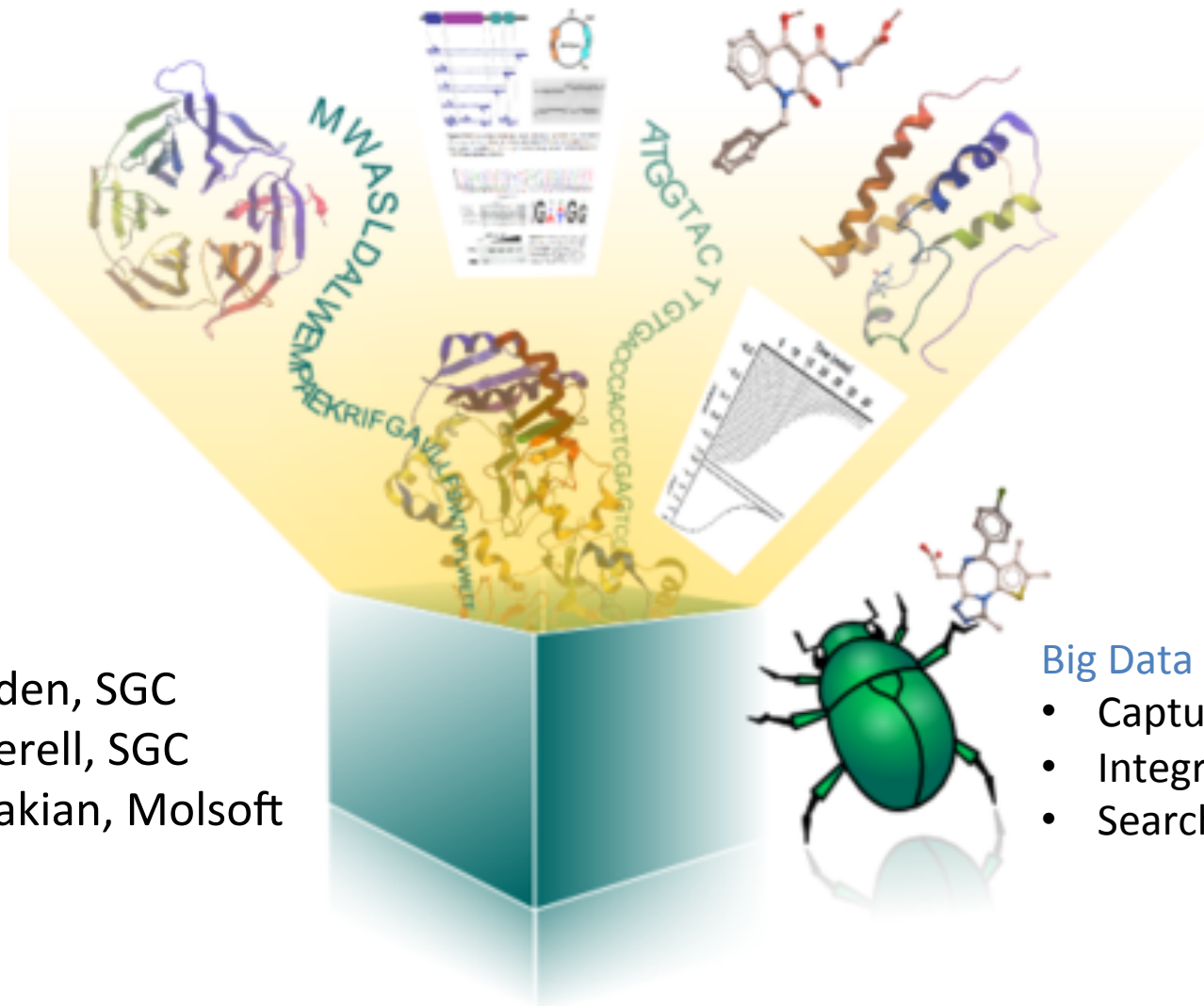
FOCUS — Development of a Global Communication and Modeling Platform for Applied and Computational Medicinal Chemists

Nikolaus Stiefl,^{*,†} Peter Gedeck,[‡] Donovan Chin,[§] Peter Hunt,^{||} Mika Lindvall,[⊥] Katrin Spiegel,^{||} Clayton Springer,[§] Scott Biller,[§] Christoph Buenemann,[#] Takanori Kanazawa,[∇] Mitsunori Kato,^{§,∇} Richard Lewis,[†] Eric Martin,[⊥] Valery Polyakov,[⊥] Ruben Tommasi,[§] John van Drie,[§] Brian Vash,[§] Lewis Whitehead,[§] Yongjin Xu,[⊥] Ruben Abagyan,[○] Eugene Raush,[○] and Max Totrov[○]

specifically Kenji Namoto, David Wayne Hughes, Paul Butler, Gregory Landrum, Sandra Mueller, Joerg Muehlbacher, Finton Sirockin, Simona Cotesta, Marc Litherland, Rajeshri Karki, Yipin Lu, Robert Pulz, Catherine Leblanc, Sabina Pecchi, Gabriel Gamber, Jay Knowles, Michael Dechantstreiter, and Thomas Dice. We also thank Jose Duca for supporting the project during the last three years. On the Molsoft side, we thank Andrew Orry, Crystal Crawford, Levon Budagyan, Polo Lam, Irina Kufareva, and Elena Arnauto^y



Scarab



Brian Marsden, SGC
David Damerell, SGC
Arman Sahakian, Molsoft

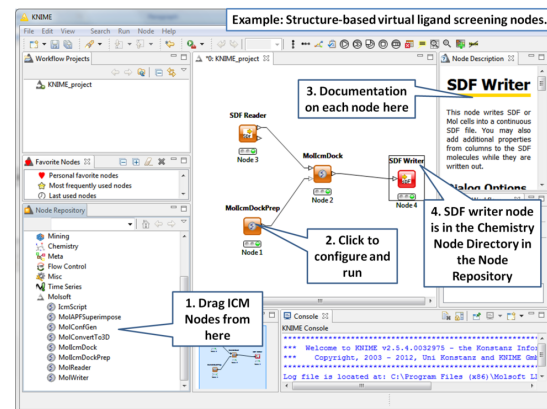
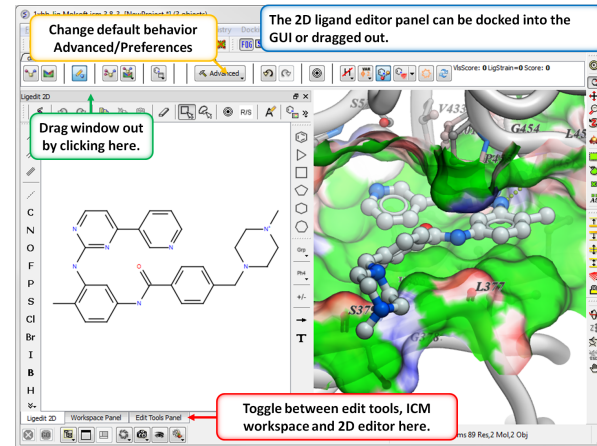
Big Data ☺ :

- Capture and Create
- Integrate
- Search and retrieve

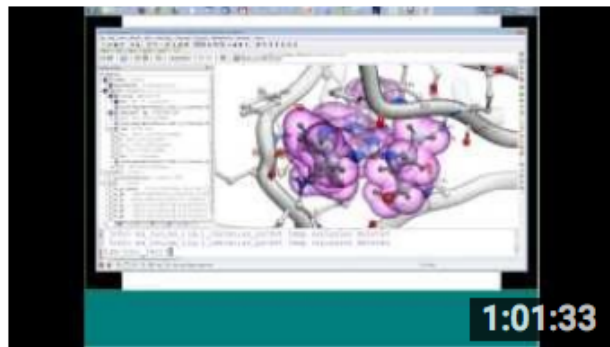


Three in One

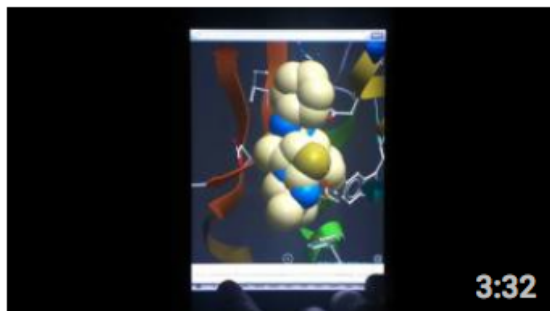
- Workstation GUI Client (eg ICM-pro, Focus, Scarab)
- ICM based backend units (models, screens, homology)
- Mobile or Web dissemination



Training Videos



MolSoft Webinar: Ligand Design using ICM 3D Interactive Ligand...
1,695 views • 2 years ago



iMolview
3,743 views • 4 years ago

MolSoft Molecules in Silico

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ICM 3D Ligand Editor: Distance Restraints and Tethers in Docking 17 views • 3 weeks ago	Ligand Editor - Ligand editing in 2D and commit changes to 3D. 6 views • 3 weeks ago	Explicit / Hybrid Docking: Fully flexible side-chains in the ICM... 17 views • 3 weeks ago	Webinar: Linking Protein Structure, Sequences and... 161 views • 1 month ago	Webinar: Molecular Graphics, Movies, and Fully Interactive 3... 293 views • 2 months ago
Webinar: Ligand Design using the ICM 3D Fully Interactive Ligand... 223 views • 2 months ago	ICM Chemist Webinar - July 2015 176 views • 3 months ago	Install MolSoft ICM Nodes for KNIME 88 views • 11 months ago	MolSoft MolCalcProp in KNIME 59 views • 11 months ago	4D Docking in the Ligand Editor. 169 views • 1 year ago
ICM Molecular Editor Tutorial 166 views • 1 year ago	MolSoft - Fully Interactive 3D Ligand Editor v3.8 254 views • 1 year ago	How to re-dock a ligand using MolSoft's ICM-Pro desktop... 457 views • 1 year ago	Introducing MolScreen from MolSoft. 49 views • 1 year ago	Covalent Docking Tutorial in ICM-Pro 486 views • 1 year ago
Protein Structure Modeling and Analysis Webinar 1,872 views • 1 year ago	Linking Protein Sequence to 3D Structure using the ICM... 778 views • 1 year ago	Introducing ICM version 3.8 563 views • 1 year ago	Molecular Graphics, Movies, and Fully Interactive 3D Documents... 964 views • 1 year ago	How to make a movie from a series of slides. 69 views • 1 year ago
Alignment Annotation in ICM 76 views • 1 year ago	Highlighting Sequence Identity in the Ligand Binding Pocket usin... 140 views • 1 year ago	Editing an Alignment in ICM 80 views • 1 year ago	Making Sequence Alignments in ICM - Linking Sequence and... 301 views • 1 year ago	How to dock to two molecules. 174 views • 1 year ago
MolSoft Webinar: Ligand Based Lead Discovery using Atomic... 488 views • 2 years ago	Webinar: Structure Based Lead Discovery using ICM Virtual... 1,213 views • 2 years ago	MolSoft Webinar: Ligand Design using ICM 3D Interactive Ligan... 1,695 views • 2 years ago	iMolview 3,743 views • 4 years ago	ICM-Browser & ActiveICM 698 views • 4 years ago

Ligand Editor



1. Select edit tools panel

2. Select impose positional restraints button

3. Select one or more atoms

4. Table values can be edited

Impose Atom Tethers

Project Summary
Lig: biotin; Rec: rec_rec1;

use	L	type	atom1	atom2	max dist	min dist	weight	rm
1	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	position	a_biotin.biotin/300/n1	35.31,10.86,-4.22	0	0	10 delete
2	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	position	a_biotin.biotin/300/n2	34.88,12.57,-5.55	0	0	10 delete
3	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	position	a_biotin.biotin/300/c5	34.28,10.36,-5.09	0	0	10 delete
4	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	position	a_biotin.biotin/300/c4	34.09,11.48,-6.11	0	0	10 delete

Results table will be displayed.

1. Table ranked by strain

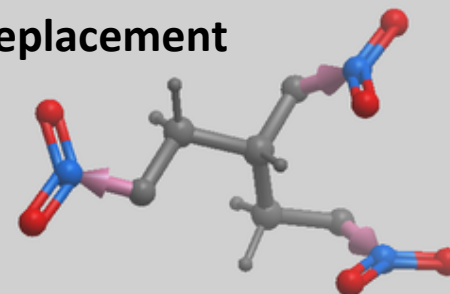
2. Display and undisplay

3. Hit is displayed

mol	Score2	Strain	L	rm	3D	name
3193	30	10	10	3.003275	3D	3-Carboxy-4,N,N-Dimethyl-2-(Octanoyloxy)Propan-Aminium

Fragment Linking

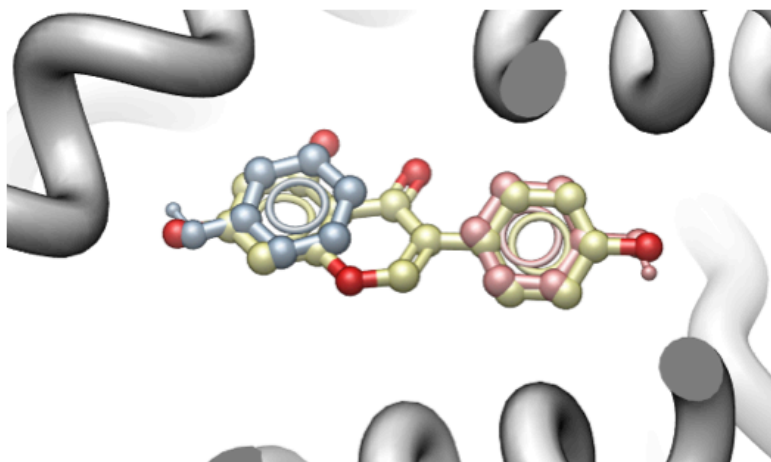
Three Point Core Replacement



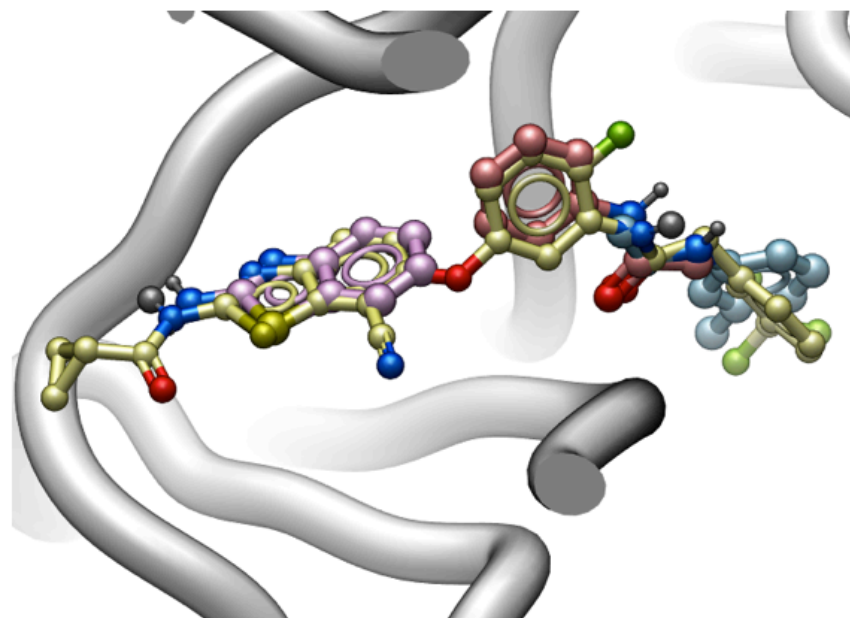
Computational Fragment Screen:



Examples of high-confidence fragment poses versus real ligands



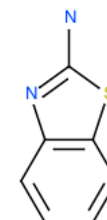
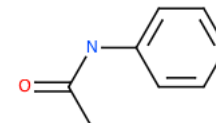
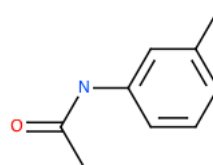
Bluish white -18.8 Red -22.7
 $p = 0.055$ $p = 0.011$



Bluish white Purple Red
 $p = 0.2$ $p = 0.047$ $p = <0.005$

Phenol fragments in PDB 1QKM (ER)

PDB 4KSP (b-RAF kinase)



Covalent Inhibitors: screening & design

Define Covalent Ligand Modification

Docking Project
covDock8

Modified Residue: Graphical Selection (1 res) ← **1. Select residue to be modified**

Reaction: Covalent_Mechaniz... ← **2. Load reactions**

Use All Reactions

Load Default Reactions

the reaction(s) that our screening ligands

Beta-lactam/Serine/PBP, beta-lactamases

Ok Cancel

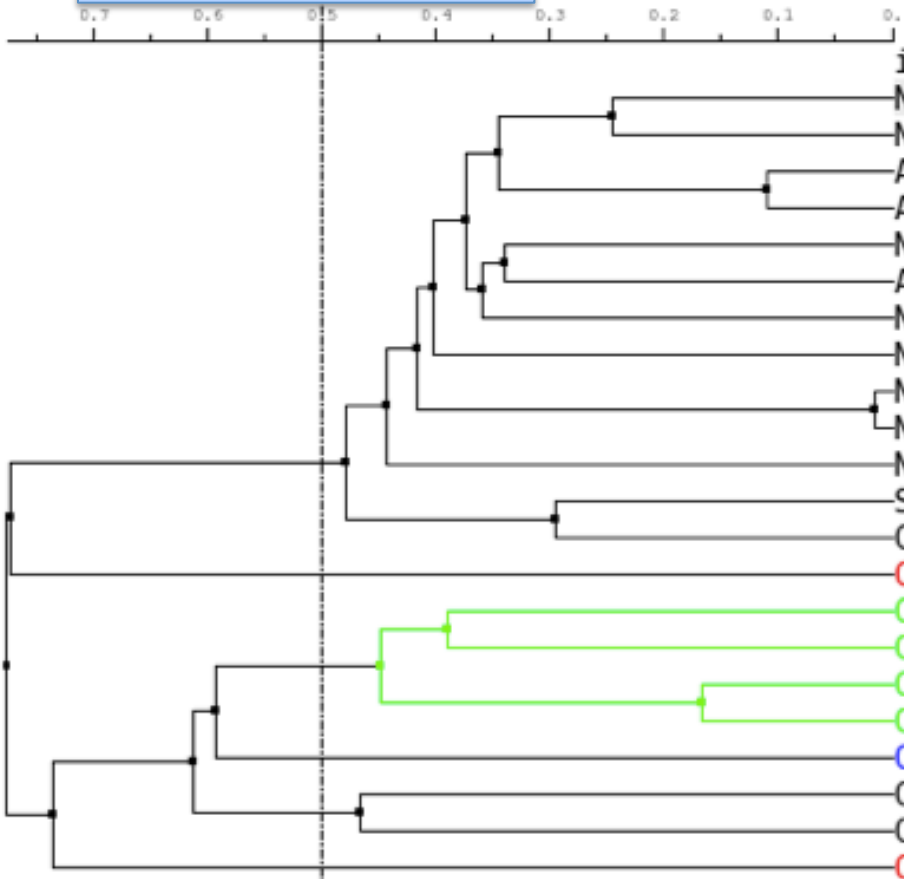
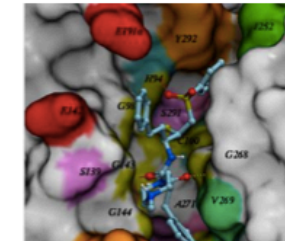
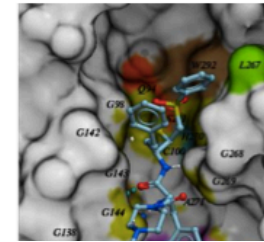
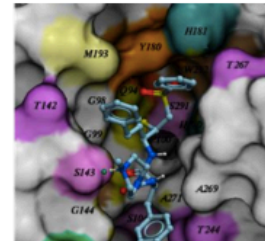
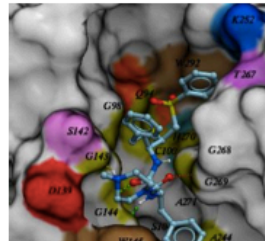
Protease with a covalent inhibitor screened by ICM

From Pocket Analysis to 3D Models

Cys-proteases from parasitic worms



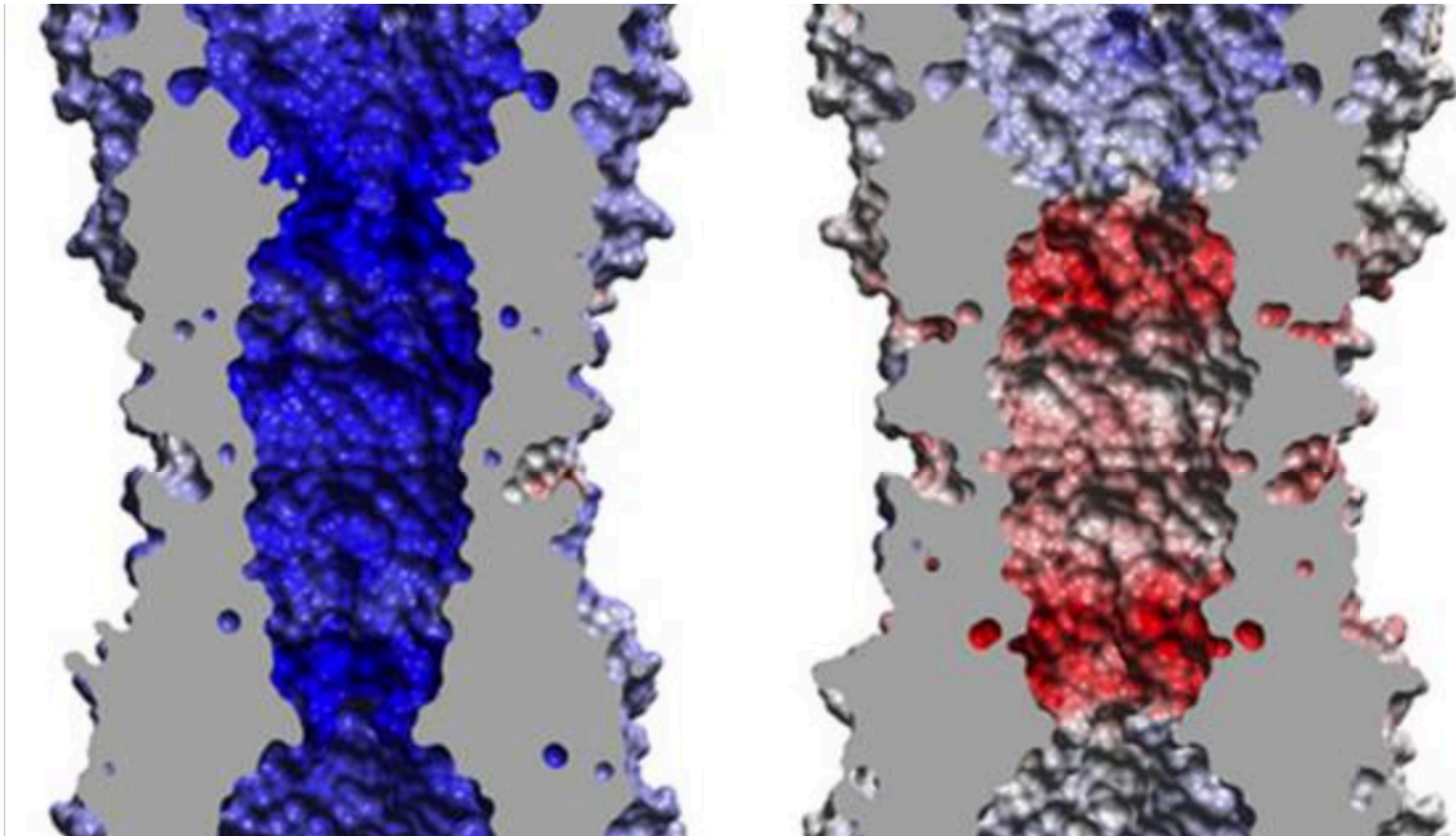
Kevin Widmer, Basel,
 Masterarbeit, 2016
 Collaboration with
 Conor Caffrey, UCSD



Protein	nSeq
id=37 nSeq=22	
Necpain	94
Na CP6	102
Acan CP1	111
Acey CP1	110
Na CP5	111
Acey CP2	116
Na CP2	116
Na CP3	112
Na CP4b	110
Na CP4	110
Na CP7	100
Sman CP	94
CathB	102
CathZ	83
CathS	133
CathK	133
CathL2	132
CathL1	132
CathH	135
Cruzipain	19
CathF	289
CathC	252

Q	C	.	.	#	.	.	.	#	#.	.	.H.	N	W	.	
Q	C	D	S	WP	NH	S	Q	A	V	FK	T	GHA	N	W	M
Q	C	S	E	YP	FK	T	Q	A	T	FK	T	GHA	N	W	M
Q	C	Y	Q	WP	HH	L	V	A	V	FS	T	AHA	N	W	Q
Q	C	Y	Q	WP	QH	L	V	A	V	YS	T	AHA	N	W	Q
Q	C	M	D	RP	RH	L	V	S	V	FA	N	AHA	N	W	R
Q	C	F	K	YP	NH	L	V	T	V	FD	T	LHA	N	W	Q
Q	C	S	T	VP	YH	M	V	T	V	FA	T	AHA	N	W	S
Q	C	A	G	HT	DE	S	T	S	I	FG	L	GHA	N	W	K
Q	C	S	E	YP	GN	G	G	N	V	FI	I	VHA	N	Y	A
H	C	S	E	YP	GN	G	G	N	V	FI	I	VHA	N	Y	Q
Q	D	V	L	SP	QH	N	V	Q	V	FI	V	LHV	N	Y	S
Q	C	L	E	IL	HH	I	E	G	V	FL	L	GHA	N	W	E
Q	C	D	N	YP	HH	E	E	A	V	FL	M	GHA	N	W	E
Q	C	G	E	ND	--	--	S	G	A	LA	Y	NHV	N	W	H
Q	C	K	N	FM	--	--	S	G	A	FF	V	NHG	N	W	F
Q	C	D	G	YM	--	--	S	A	A	FQ	L	NHA	N	W	L
Q	C	Q	N	FM	--	--	S	A	A	FQ	L	DHG	N	W	A
Q	C	E	N	LM	--	--	S	A	A	FL	M	DHG	N	W	A
Q	C	H	Q	LP	--	--	S	A	V	FM	V	NHA	N	W	C
Q	C	S	S	LM	--	--	A	A	A	WM	L	DHG	N	W	E
Q	C	K	M	LP	--	--	S	A	A	MQ	I	DHA	N	W	M
Q	C	Q	E	FP	--	--	A	A	V	FL	L	NHA	N	W	I

Boundary Element (REBEL) electrostatics

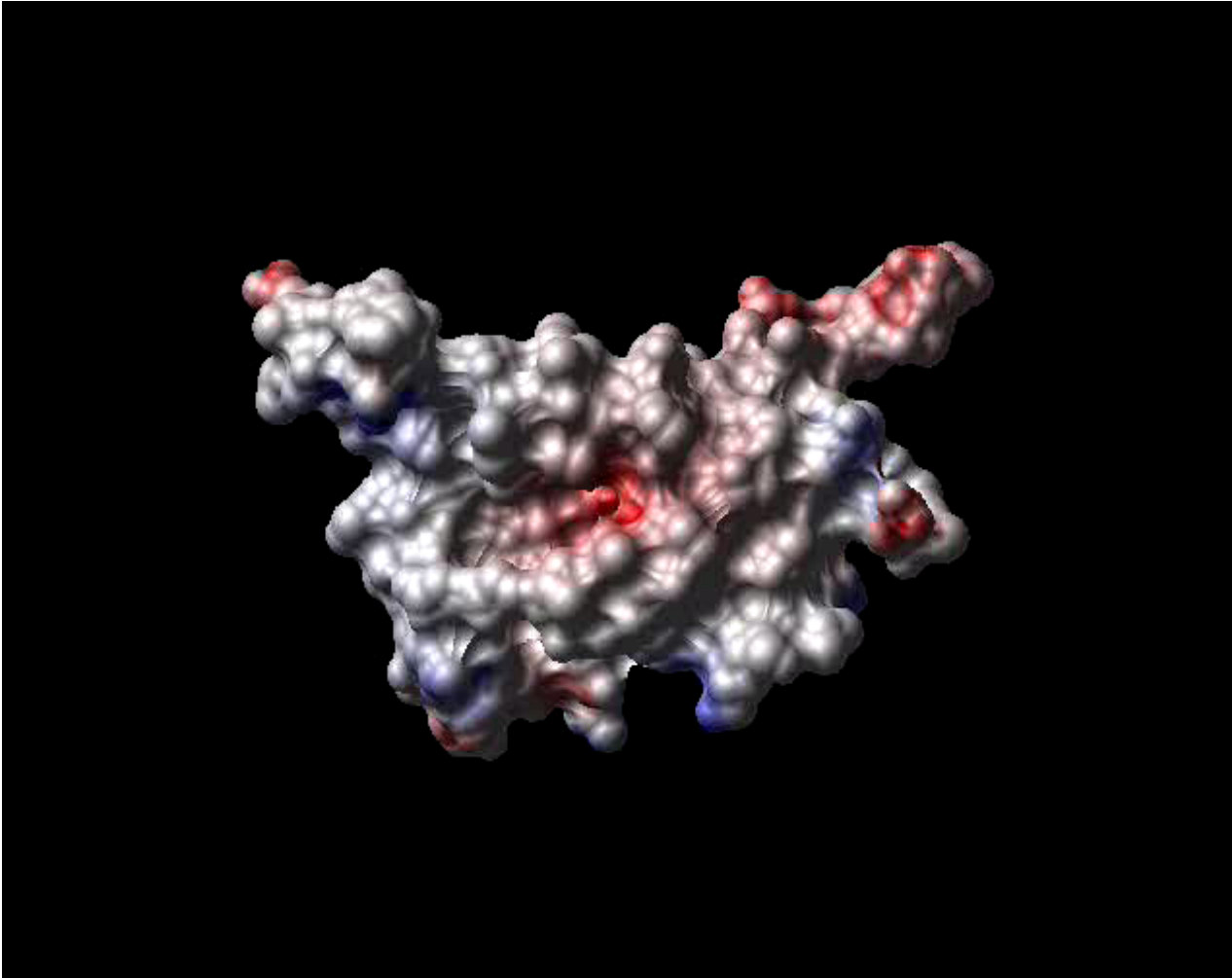


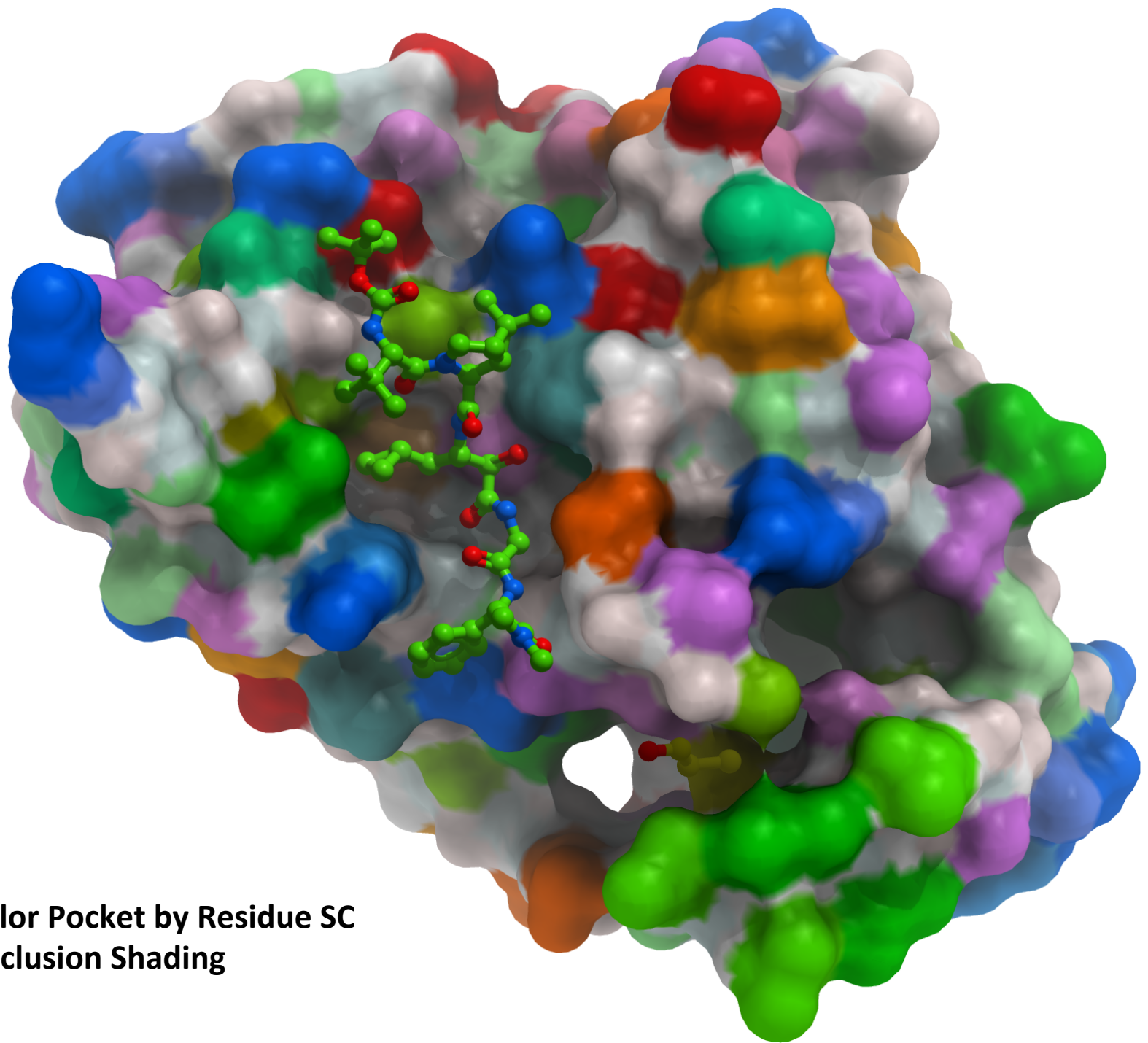
Bennett et al. **An electrostatic mechanism for Ca^{2+} -mediated regulation of gap junction channels.** *Nature Comm*, 2016

Totrov, Abagyan. **Rapid boundary element solvation electrostatics calculations in folding simulations: successful folding of a 23-residue peptide.** *Biopolymers*. 2001;60(2):124-33

ICM pocket Finder

Jianghong An, Maxim Totrov, R. Abagyan. (2005) *Pocketome: Comprehensive Identification & Classification of Ligand Binding Envelopes*, *Mol. Cell Proteomics*





- **Color Pocket by Residue SC**
- **Occlusion Shading**

IN THEORY THERE IS
NO DIFFERENCE
BETWEEN THEORY AND
PRACTICE. IN
PRACTICE THERE IS.

YOGI BERRA

Docking to the Pocketome

www.pocketome.org
Pocketome.ucsd.edu

Molecular & Cellular Proteomics

~3000 ensembles

Volume 4, Number 6, June 2005

www.mcponline.org

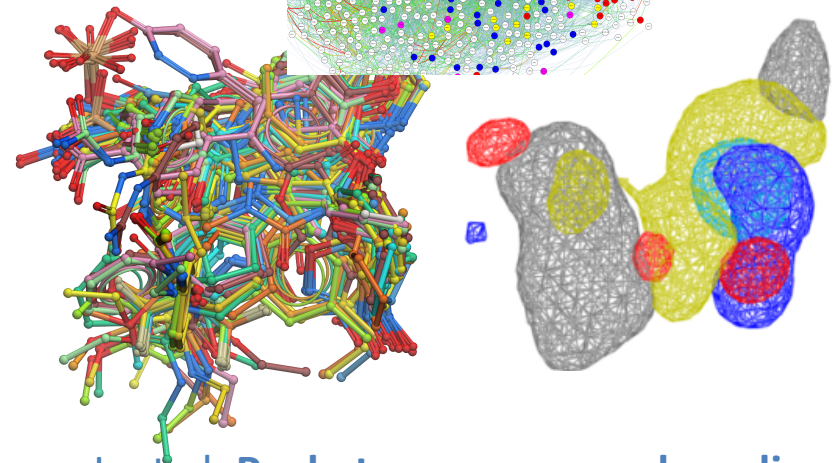
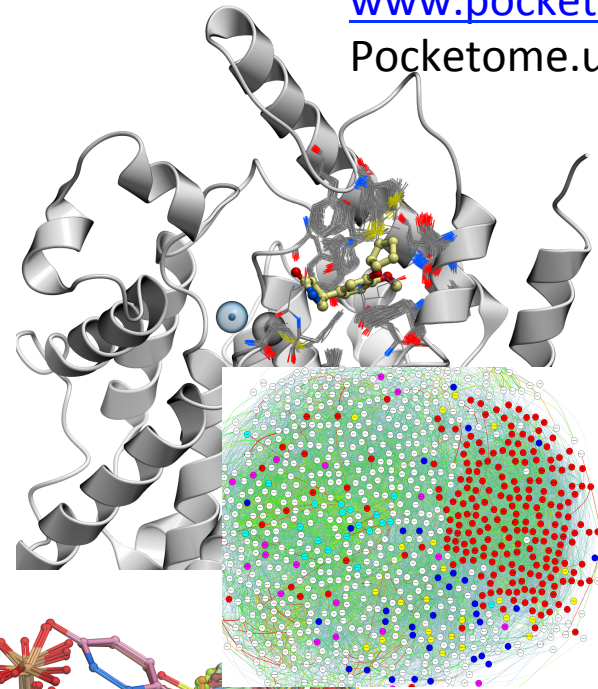


The pocketome

In this issue

- Phosphotyrosine Proteomics of IFN α Signaling
- Proteomic Analysis of Human Adult Stem Cell Adipogenesis
- Purification of Ubiquitin Conjugates and Rpn10 Substrates
- Identification and Classification of Ligand Envelopes
- PROVALT: Protein Validation Technology
- Antibody Array Normalization
- MudPIT Analysis of 14-3-3 α
- Rice Basal Region Proteome
- Phosphopeptide Methylation and Detection by MALDI Q-TOF MS
- A Yeast/Bacteria Two-hybrid System
- Metalloprotein Assay
- Improving Protein Identification Using Complementary Pairs

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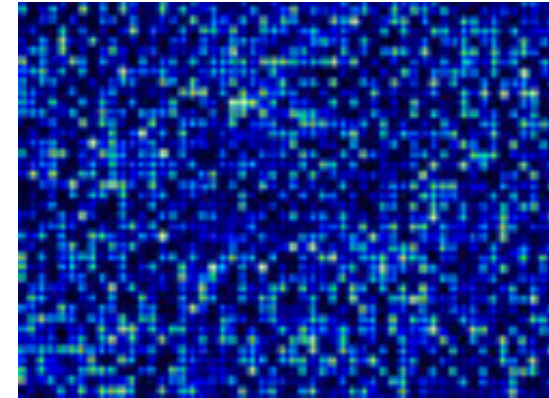
Kufareva I, et al. Pocketome: an encyclopedia of binding sites in 4D. *Nucleic Acids Res.* 2012

An, Totrov, Abagyan, 2005

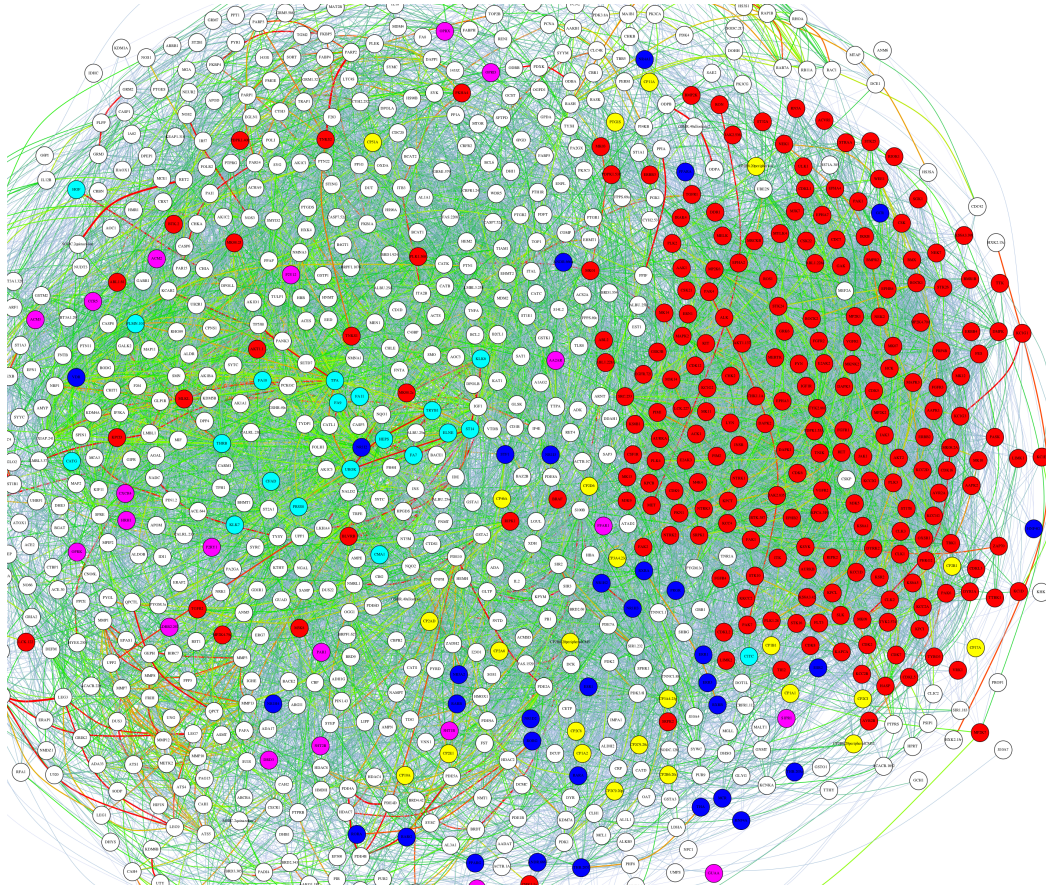
Drugs



Profiles of relevant Patient Cells



MULTIPLE Drug Targets (the Pocketome)

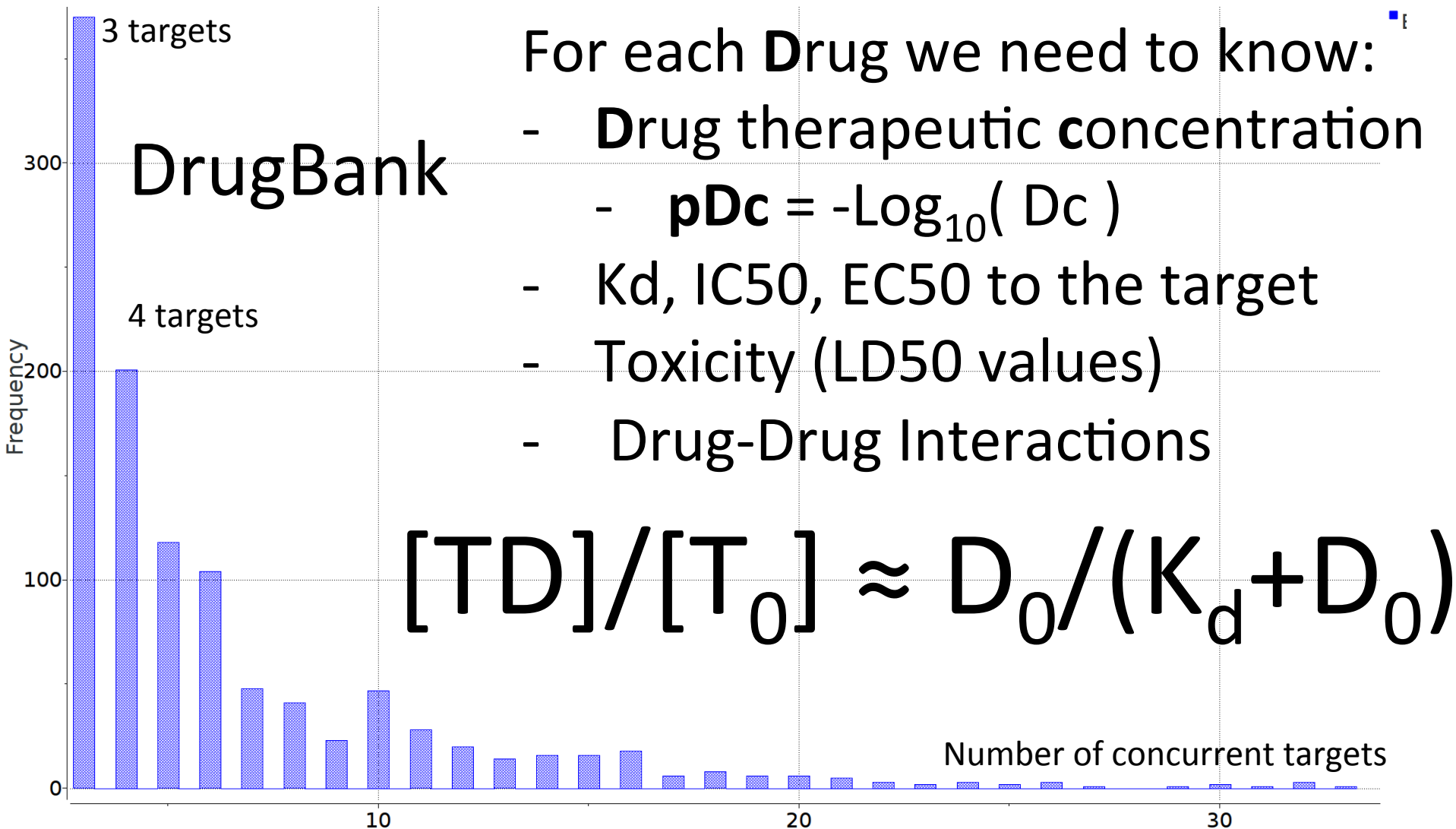


Optimal Match with one or few drugs



Substantial Multi-target Pharmacology

Targets of Drugs are under-Discovered and the binding is under-Quantified



ChEMBL Database

- Example: Acitivites of Bosutinib above the trough drug levels + a margin to achive > 75% inhibition
- pDc+0.5

chembl_bosutinib_targets

	1	2	3	4
0	uniprot_id ABL1_HUMAN fullname Tyrosine-protein kinase ABL1 pAct 10.54	uniprot_id M4K5_HUMAN fullname Mitogen-activated protein kin pAct 8.33	uniprot_id YES_HUMAN fullname Tyrosine-protein kinase Yes pAct 8.47	uniprot_id ABL2_HUMAN fullname Abelson tyrosine-protein kina pAct 8.33
4	uniprot_id LCK_HUMAN fullname Tyrosine-protein kinase Lck pAct 8.23	uniprot_id ERBB3_HUMAN fullname Receptor tyrosine-protein kin pAct 8.11	uniprot_id SRC_HUMAN fullname Proto-oncogene tyrosine-pro pAct 8	uniprot_id FGR_HUMAN fullname Tyrosine-protein kinase Fgr pAct 8.96
8	uniprot_id fullname pAct 8.96	uniprot_id GAK_HUMAN fullname Cyclin-G-associated kinase pAct 8.89	uniprot_id FRK_HUMAN fullname Tyrosine-protein kinase FRK pAct 8.85	uniprot_id FYN_HUMAN fullname Tyrosine-protein kinase Fyn pAct 8.74
12	uniprot_id STK35_HUMAN fullname Serine/threonine-protein kina pAct 8.7	uniprot_id BTK_HUMAN fullname Tyrosine-protein kinase BTK pAct 8.6	uniprot_id ACK1_HUMAN fullname Activated CDC42 kinase 1 pAct 8.57	uniprot_id M4K2_HUMAN fullname Mitogen-activated protein kin pAct 8.51
16	uniprot_id MINK1_HUMAN fullname Misshapen-like kinase 1 pAct 8.49	uniprot_id HCK_HUMAN fullname Tyrosine-protein kinase HCK pAct 8.49	uniprot_id BLK_HUMAN fullname Tyrosine-protein kinase Blk pAct 8.48	uniprot_id STK24_HUMAN fullname Serine/threonine-protein kina pAct 8.41
20	uniprot_id LYN_HUMAN fullname Tyrosine-protein kinase Lyn pAct 8.38	uniprot_id SLK_HUMAN fullname STE20-like serine/threonine-p pAct 8.33	uniprot_id M4K3_HUMAN fullname Mitogen-activated protein kin pAct 8.29	uniprot_id EPHB4_HUMAN fullname Ephrin type-B receptor 4 pAct 8.26
24	uniprot_id EPHA3_HUMAN fullname Ephrin type-A receptor 3 pAct 8.24	uniprot_id STK10_HUMAN fullname Serine/threonine-protein kina pAct 8.15	uniprot_id BLK_MOUSE fullname Tyrosine-protein kinase Blk pAct 8.14	uniprot_id BMX_HUMAN fullname Cytoplasmic tyrosine-protein pAct 8.1
28	uniprot_id MP2K5_HUMAN fullname Dual specificity mitogen-activ pAct 8.09	uniprot_id M4K4_HUMAN fullname Mitogen-activated protein kin pAct 8.09	uniprot_id EPHB2_HUMAN fullname Ephrin type-B receptor 2 pAct 8.08	uniprot_id EPHA8_HUMAN fullname Ephrin type-A receptor 8 pAct 8.05
32	uniprot_id MP2K2_HUMAN fullname Dual specificity mitogen-activ pAct 8	uniprot_id M4K1_HUMAN fullname Mitogen-activated protein kin pAct 7.82	uniprot_id M3K19_HUMAN fullname Mitogen-activated protein kin pAct 7.8	uniprot_id EGFR_HUMAN fullname Epidermal growth factor rece pAct 7.74
36	uniprot_id EPHA2_HUMAN fullname Ephrin type-A receptor 2 pAct 7.74	uniprot_id EPHA4_HUMAN fullname Ephrin type-A receptor 4 pAct 7.74	uniprot_id MP2K1_HUMAN fullname Dual specificity mitogen-activ pAct 7.72	uniprot_id NTRK1_HUMAN fullname High affinity nerve growth fac pAct 7.66
40	uniprot_id ERBB4_HUMAN fullname Receptor tyrosine-protein kin pAct 7.59	uniprot_id EPHA5_HUMAN fullname Ephrin type-A receptor 5 pAct 7.57	uniprot_id NTRK2_HUMAN fullname BDNF/NT-3 growth factors rec pAct 7.57	uniprot_id SIK2_HUMAN fullname Serine/threonine-protein kina pAct 7.54
44	uniprot_id M3K2_HUMAN fullname Mitogen-activated protein kin pAct 7.52	uniprot_id SIK1_HUMAN fullname Serine/threonine-protein kina pAct 7.52	uniprot_id TNK1_HUMAN fullname TRAF2 and NCK-interacting pr pAct 7.51	uniprot_id CSK_HUMAN fullname Tyrosine-protein kinase CSK pAct 7.49
48	uniprot_id EPHB1_HUMAN fullname Ephrin type-B receptor 1 pAct 7.48	uniprot_id MST4_HUMAN STK26_HUMAN fullname Serine/threonine-protein kina pAct 7.43	uniprot_id STK33_HUMAN fullname Serine/threonine-protein kina pAct 7.43	uniprot_id TXK_HUMAN fullname Tyrosine-protein kinase TXK pAct 7.4
52	uniprot_id IKKε_HUMAN fullname Inhibitor of nuclear factor kap pAct 7.28	uniprot_id UFE1L_HUMAN fullname Tyrosine-protein kinase rece pAct 7.28	uniprot_id KSYK_HUMAN fullname Tyrosine-protein kinase SYK pAct 7.28	uniprot_id M3K3_HUMAN fullname Mitogen-activated protein kin pAct 7.27
56	uniprot_id TYRO3_HUMAN fullname Tyrosine-protein kinase recep pAct 7.21	uniprot_id SIK3_HUMAN fullname Serine/threonine-protein kina pAct 7.19	uniprot_id KIT_HUMAN fullname Mast/stem cell growth factor pAct 7.14	uniprot_id MLTK_HUMAN fullname Mitogen-activated protein kin pAct 7.1
60	uniprot_id DMPK_HUMAN fullname Myotinin-protein kinase pAct 7.04	uniprot_id KCC1D_HUMAN fullname Calcium/calmodulin-depender pAct 7.04	uniprot_id WEE2_HUMAN fullname Wee1-like protein kinase 2 pAct 7.03	uniprot_id SRMS_HUMAN fullname Tyrosine-protein kinase Srms pAct 7
64	uniprot_id M3K7_HUMAN fullname Mitogen-activated protein kin pAct 7	uniprot_id M4K4_HUMAN fullname Mitogen-activated protein kin pAct 6.96	uniprot_id KC1E_HUMAN fullname Casein kinase I isoform epsil pAct 6.96	uniprot_id MERTK_HUMAN fullname Tyrosine-protein kinase Mer pAct 6.96
68	uniprot_id DDR1_HUMAN fullname Epithelial discoidin domain-co pAct 6.92	uniprot_id CHK2_HUMAN fullname Serine/threonine-protein kina pAct 6.9	uniprot_id HIPK4_HUMAN fullname Homeodomain-interacting pro pAct 6.88	uniprot_id FAK2_HUMAN fullname Protein-tyrosine kinase 2-bet pAct 6.87
72	uniprot_id DDR2_HUMAN fullname Discoidin domain-containing r pAct 6.85	uniprot_id NF1_HUMAN fullname Nucleophosmin pAct 6.82	uniprot_id M3K7_HUMAN fullname Serine/threonine-protein kina pAct 6.77	uniprot_id KCC2G_HUMAN fullname Calcium/calmodulin-depender pAct 6.74
76	uniprot_id STK4_HUMAN fullname Serine/threonine-protein kina pAct 6.72	uniprot_id PGFRB_HUMAN fullname Platelet-derived growth fact pAct 6.7	uniprot_id EPHB3_HUMAN fullname Ephrin type-B receptor 3 pAct 6.68	uniprot_id NUAK2_HUMAN fullname NUAK family SNF1-like kinase pAct 6.66
80	uniprot_id KC1D_HUMAN fullname Casein kinase I isoform delta pAct 6.62	uniprot_id TBK1_HUMAN fullname Serine/threonine-protein kina pAct 6.6	uniprot_id E2AK4_HUMAN fullname p15-2 alpha kinase GCN2 (EC pAct 6.55	uniprot_id KC1A_HUMAN fullname Casein kinase I isoform alpha pAct 6.55
84	uniprot_id TEC_HUMAN fullname Tyrosine-protein kinase Tec pAct 6.55	uniprot_id CLK3_HUMAN fullname Dual specificity protein kinase pAct 6.52	uniprot_id FES_HUMAN fullname Tyrosine-protein kinase Fes/Fr pAct 6.48	uniprot_id PHKG1_HUMAN fullname Phosphorylase b kinase gam pAct 6.47
88	uniprot_id TAOK3_HUMAN fullname Serine/threonine-protein kina pAct 6.47	uniprot_id MAST1_HUMAN fullname Microtubule-associated serin pAct 6.46	uniprot_id BMP2K_HUMAN fullname BMP-2-inducible protein kinas pAct 6.46	uniprot_id PMYT1_HUMAN fullname Membrane-associated tyrosin pAct 6.46
92	uniprot_id FER_HUMAN fullname Tyrosine-protein kinase Fer pAct 6.44	uniprot_id IRAK4_HUMAN fullname Interleukin-1 receptor-associ pAct 6.44	uniprot_id ROCK2_HUMAN fullname Rho-associated protein kinas pAct 6.44	uniprot_id STK3_HUMAN fullname Serine/threonine-protein kina pAct 6.43
96	uniprot_id DUSTY_HUMAN fullname Dual serine/threonine and tyr pAct 6.42	uniprot_id CSF1R_HUMAN fullname Macrophage colony-stimulati pAct 6.42	uniprot_id ULK2_HUMAN fullname Serine/threonine-protein kina pAct 6.35	uniprot_id ULK3_HUMAN fullname Serine/threonine-protein kina pAct 6.34
100	uniprot_id M3K13_HUMAN fullname Mitogen-activated protein kin pAct 6.32	uniprot_id MYLK_HUMAN fullname Myosin light chain kinase, smc pAct 6.31	uniprot_id LRRK2_HUMAN fullname Leucine-rich repeat serine/thr pAct 6.3	uniprot_id WEE1_HUMAN fullname Wee1-like protein kinase pAct 6.29

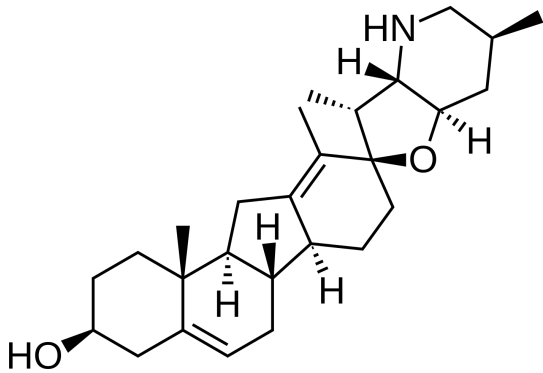
Multi-target pharmacology: friend of foe?

Discovery of *Useful* Additional Activities of Existing Drugs

MTP opportunities

- Better drugs for a specific target
- Additional targets for specific drugs
- Targets for a drug with unknown mechanism of action

Dysregulated Hedgehog Pathway. Smoothened Receptor

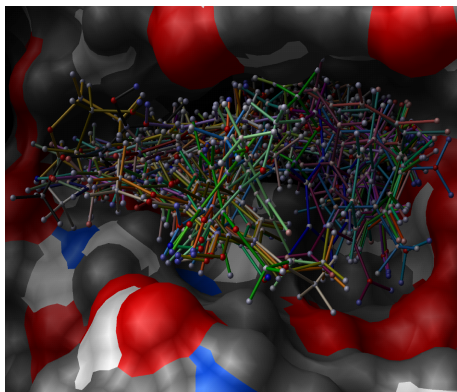


Cyclopamine: teratogen from
Corn Lily.

Hh-pathway: embryonic
development, differentiation,
cancer



Converting Pocket Ensembles with Co-crystallized Ligands into Docking/Binding Models



From **Score** to **pKd** or ΔG ?

- Target specific Score shifts
- Re-trained docking Score
- Full pKd training on docked poses

2008, Kufareva et al., *JMC*, Profiling Kinases

2010, Park et al. *JCAMD*, 17 Nuclear Receptors

2014, Chen et al., *FMC*, 37 Pocket/Ligand Ensembles

2016, Lam et al., (MolScreen), >3000 Models

- Screening for a real multi-target profile of drugs
- Repurposing drugs or reviving abandoned candidates
- Predicting targets of hits from phenotypic assays
- Predicting adverse effects of drugs and environmental chemicals, additives and metabolites

$$\Delta G_l \approx \sum_i \sum_k \omega_i^k E_i^{kl}$$

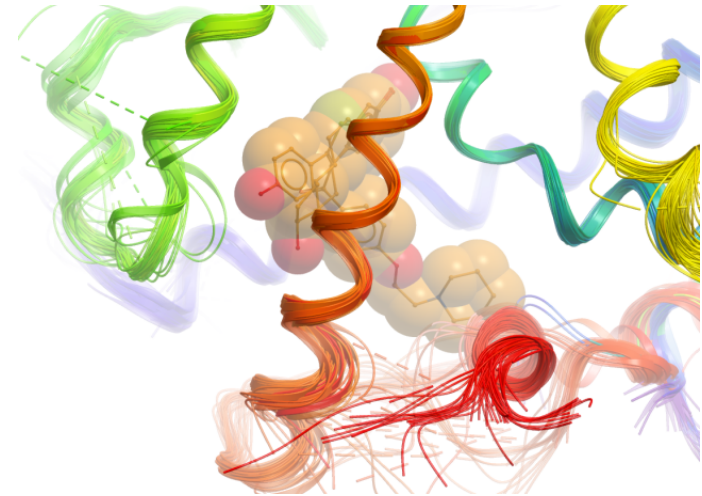
Pocket Selection: Rueda, et al. 2012, Alibero, *JCIM*

Bottegoni G, Rocchia W, Rueda M, Abagyan R, Cavalli A. **Systematic exploitation of multiple receptor conformations for virtual ligand screening.** *PLoS One*. 2011

Pocketome-derived Target Screen: Docking-Model Types and Outputs

DPC: Docking to Pocket, (343)

- Multi-conformational, 4D
- Template assisted, DP
- Selection, *single* cluster
- Pose + Class plus Activity (pIC50)



DFZ: Docking to Ligand (504)

Fields, Z-Score (normalized docking Score)

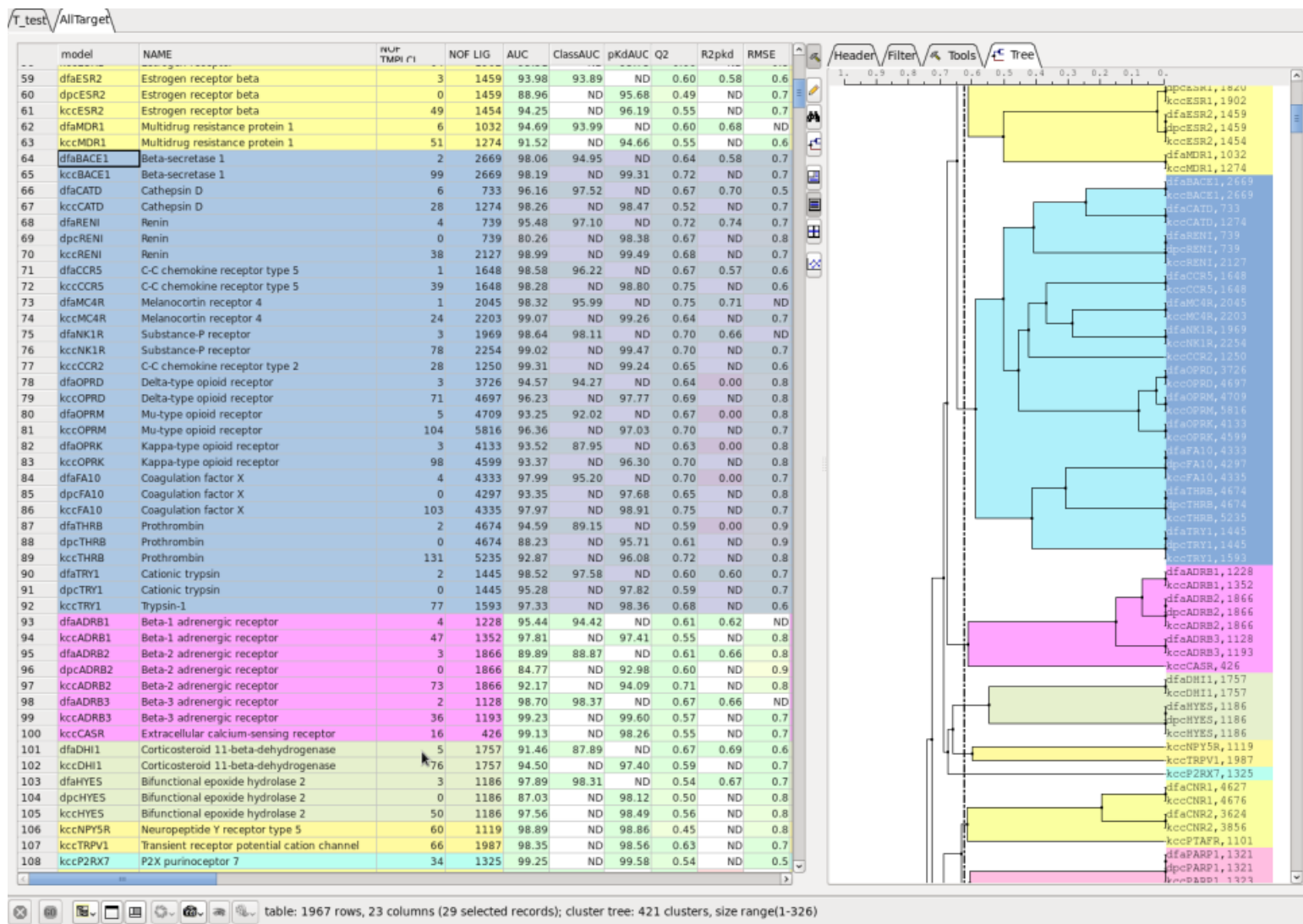
KCC: 2D Random Forest: (1139)

DFA: Docking to Ligand Fields+Act training (1035)

- Pocket driven or APF-Superposition
- *Multiple* Clusters
- Pose + Activity (pP, pIC50)

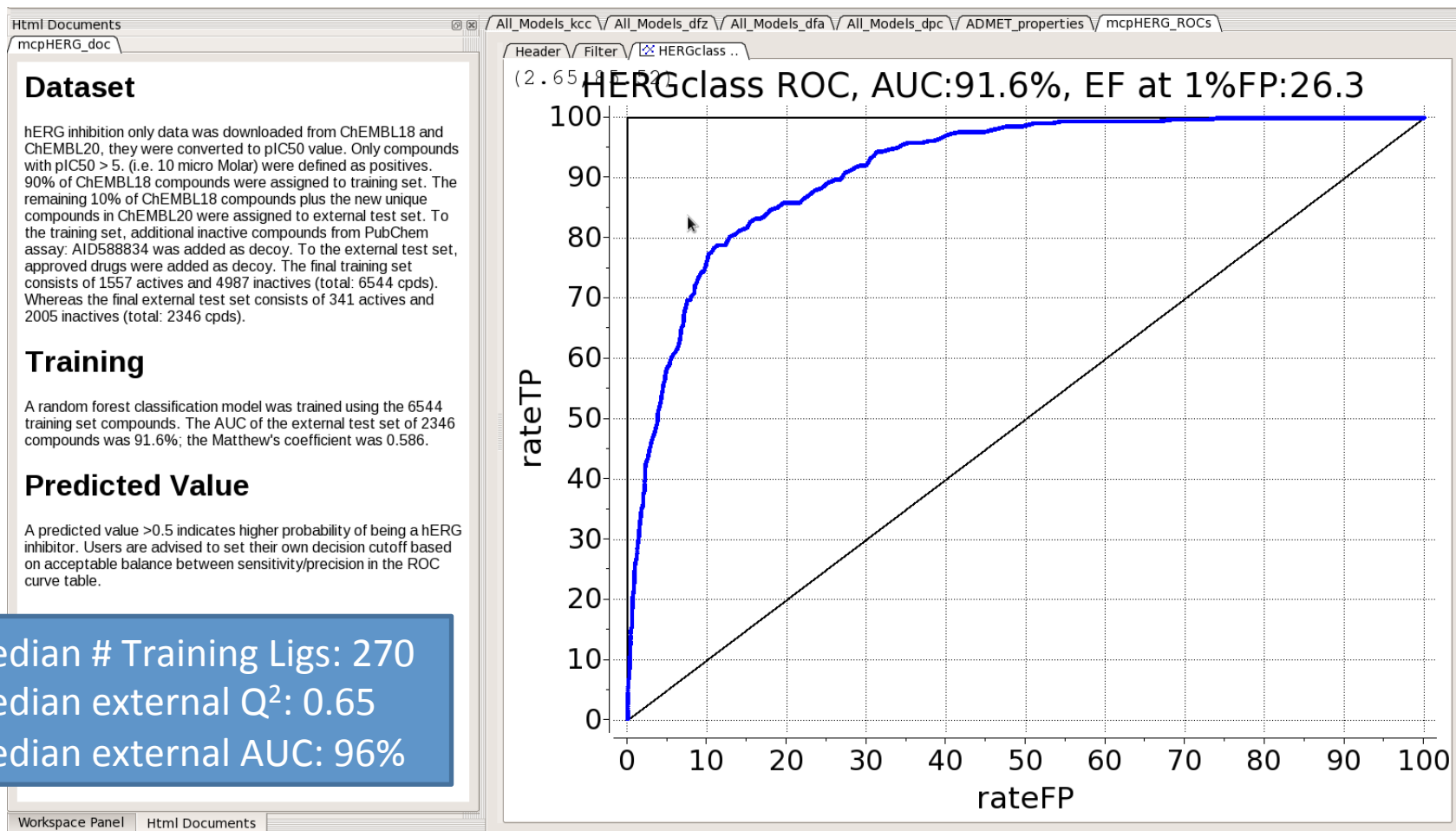


~3000 MolScreen Models clustered

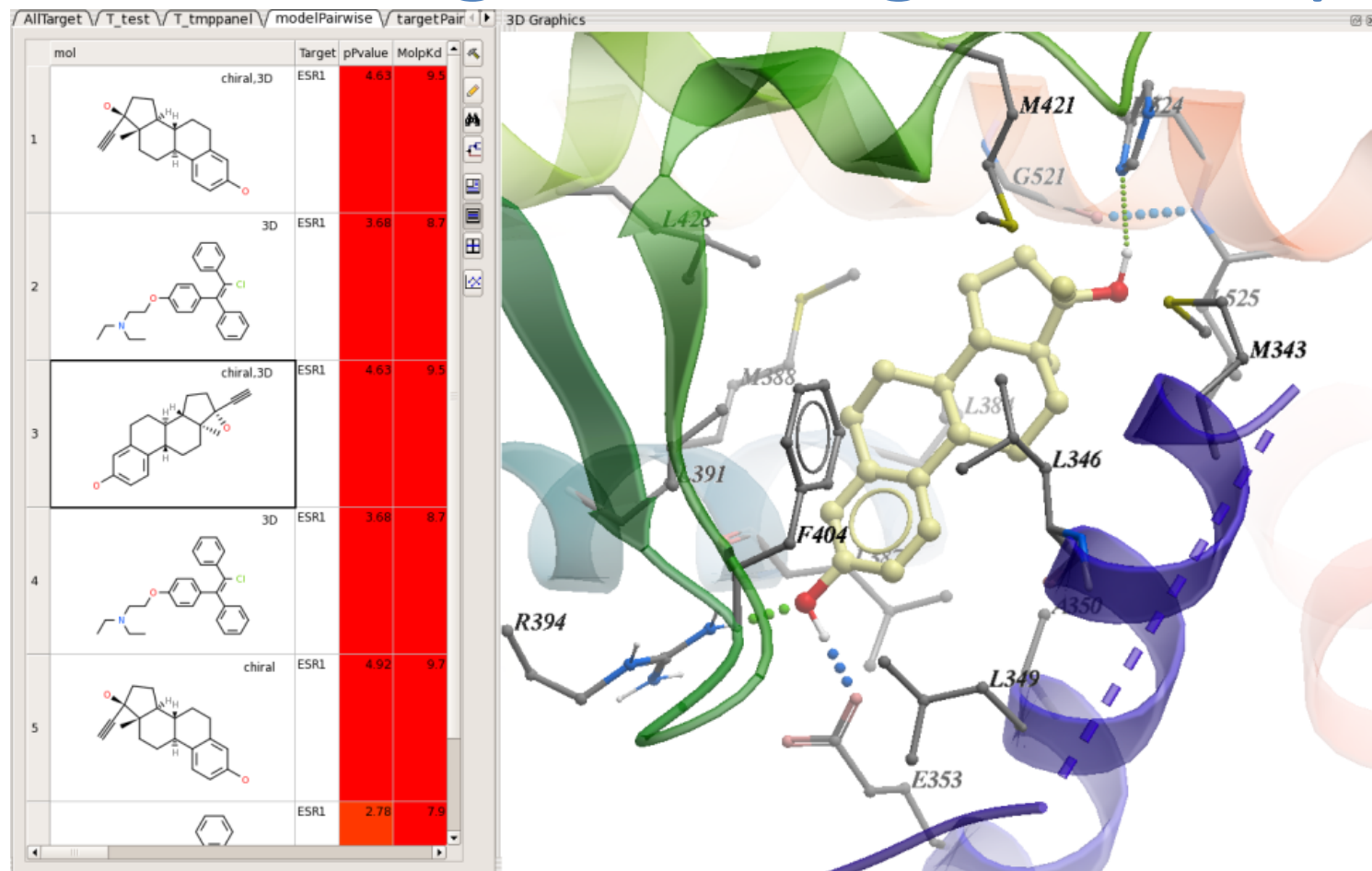


Individual models: performance and training details

- Load Panel -> Check Model Performance:



Best Docking Pose to go with c-pKd

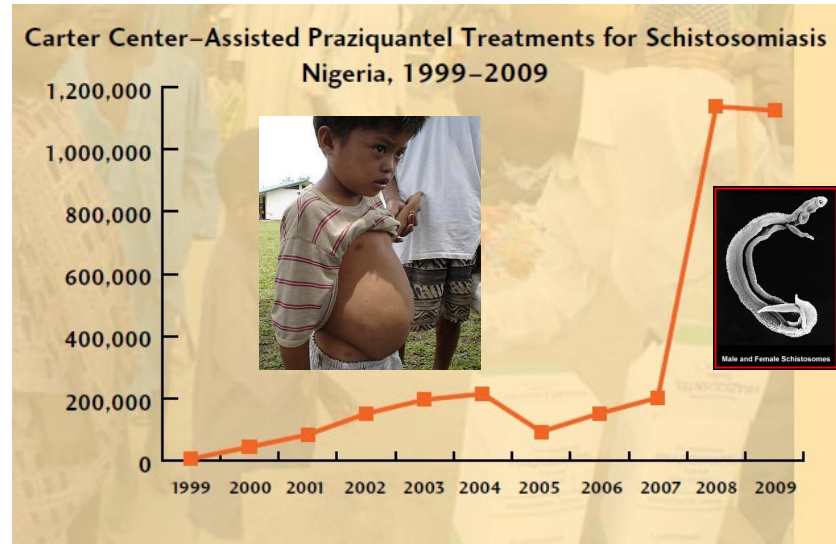
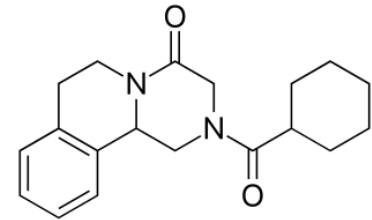


- dfa, dpc, dfz models are based on docking
- pKd prediction based on random forest training if 3D methods fail or are not accurate enough

Discovering MOA with the Pocketome

Parasitic flatworms and Praziquantel

- *Schistosomiasis*: 200-400M
 - Some develop CNS symptoms
- *Hydatid* disease (Echinococcus)
- *Cysticercosis*: brain/muscle by eggs and larvae of the pork tapeworm
- Praziquantel against tapeworms and flukes (schisto: single dose)
- PZQ is extremely well tolerated. But .. The **MOA is unknown** and resistance is imminent



Praziquantel target screen

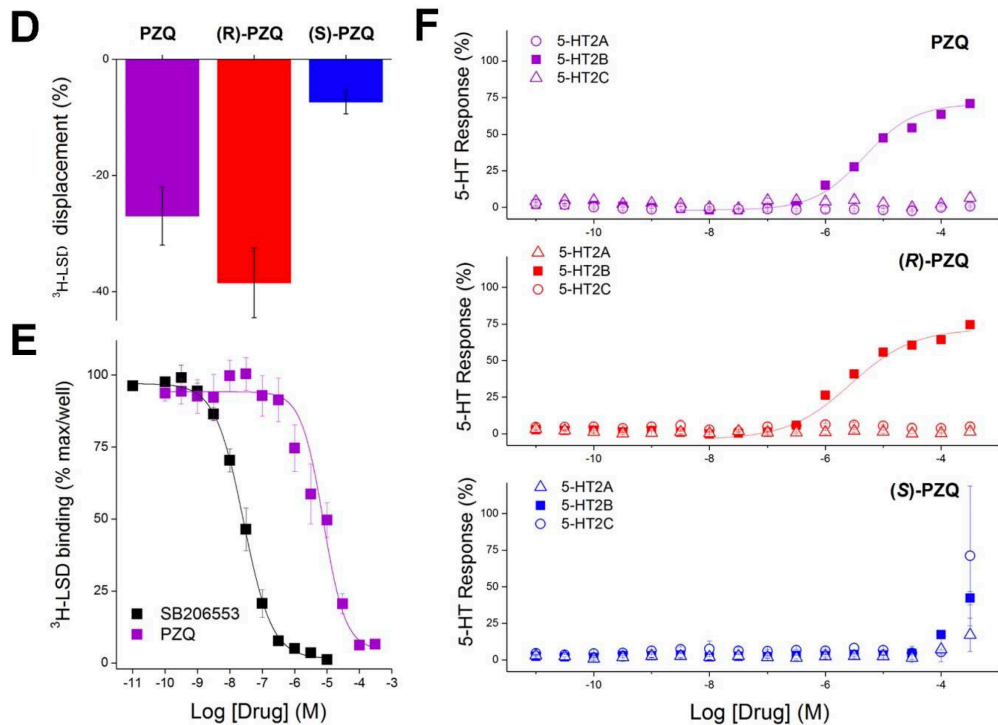
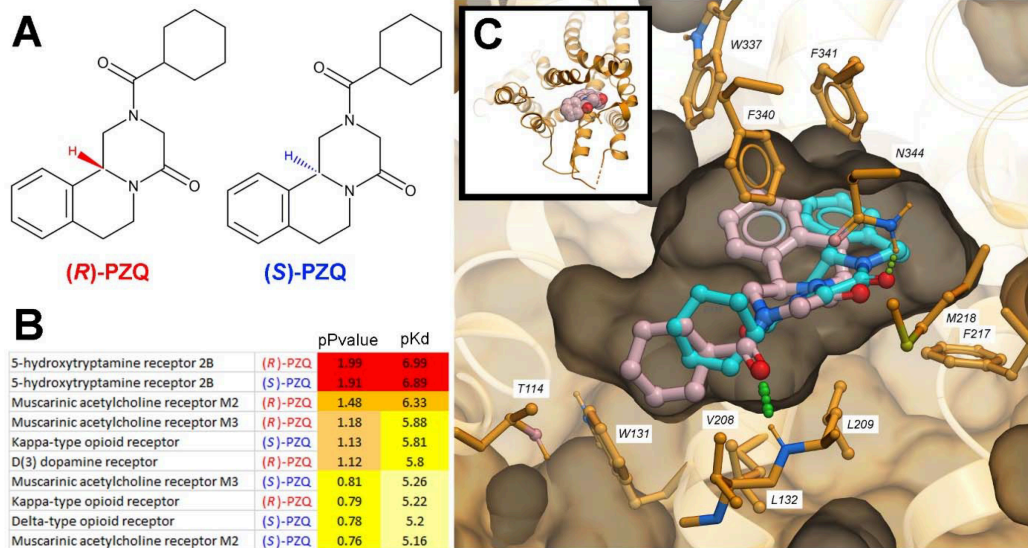
Collaboration with
Pauline Cupid, Brian Roth
Charles Cunningham,
Jonathan Marchant
Nature Comm. (submitted)

Docking PZQ-R/S to 343 4D pockets

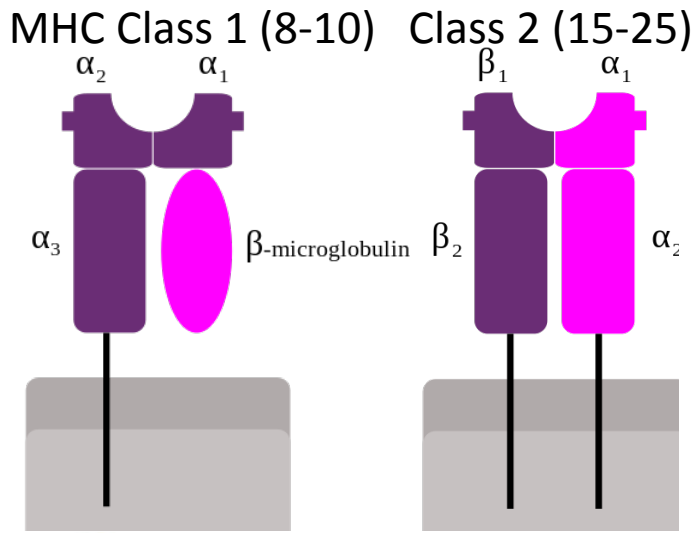
Target	Code	pPvalue	Mol_pKd	pKd_Error	Drug
5-hydroxytryptamine receptor 2B	5HT2B	1.99	6.99	1.44	(R)-PZQ
5-hydroxytryptamine receptor 2B	5HT2B	1.91	6.89	1.44	(S)-PZQ
Muscarinic acetylcholine receptor M2	ACM2	1.48	6.33	1.24	(R)-PZQ
Muscarinic acetylcholine receptor M3	ACM3	1.18	5.88	1.20	(R)-PZQ
Kappa-type opioid receptor	OPRK	1.13	5.81	1.28	(S)-PZQ
D(3) dopamine receptor	DRD3	1.12	5.80	1.28	(R)-PZQ
Muscarinic acetylcholine receptor M3	ACM3	0.81	5.26	1.20	(S)-PZQ
Kappa-type opioid receptor	OPRK	0.79	5.22	1.28	(R)-PZQ
Delta-type opioid receptor	OPRD	0.78	5.20	1.23	(S)-PZQ
Muscarinic acetylcholine receptor M2	ACM2	0.76	5.16	1.24	(S)-PZQ
Adenosine receptor A2a	AA2AR	0.45	4.25	1.26	(R)-PZQ
Adenosine receptor A2a	AA2AR	0.48	3.54	1.26	(S)-PZQ
Delta-type opioid receptor	OPRD	0.39	3.44	1.23	(R)-PZQ
Beta-2 adrenergic receptor	ADRB2	0.82	2.41	1.27	(R)-PZQ
Beta-2 adrenergic receptor	ADRB2	0.23	ND	ND	(S)-PZQ
D(3) dopamine receptor	DRD3	0.05	ND	ND	(S)-PZQ
Corticotropin-releasing factor receptor 1	CRFR1	0.55	ND	ND	(S)-PZQ
Corticotropin-releasing factor receptor 1	CRFR1	0.68	ND	ND	(R)-PZQ

Figure 1

Predicting a human target of the schistosomal drug Praziquantel

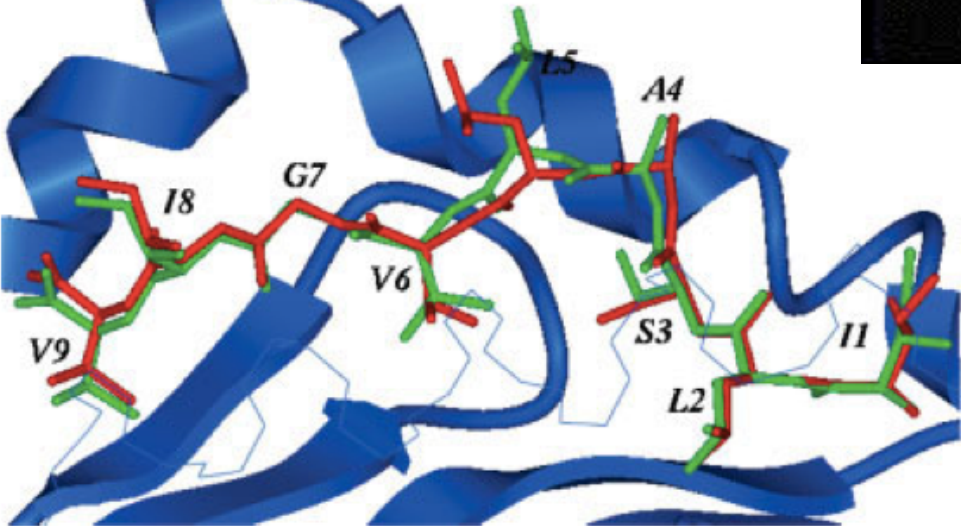


Collaboration with
 Pauline Cupid, Brian Roth
 Charles Cunningham,
 Jonathan Marchant
Nature Comm. (submitted)



Docking flexible phosphorylated peptide to a receptor
(pYLRVA to V-SRC SH2)

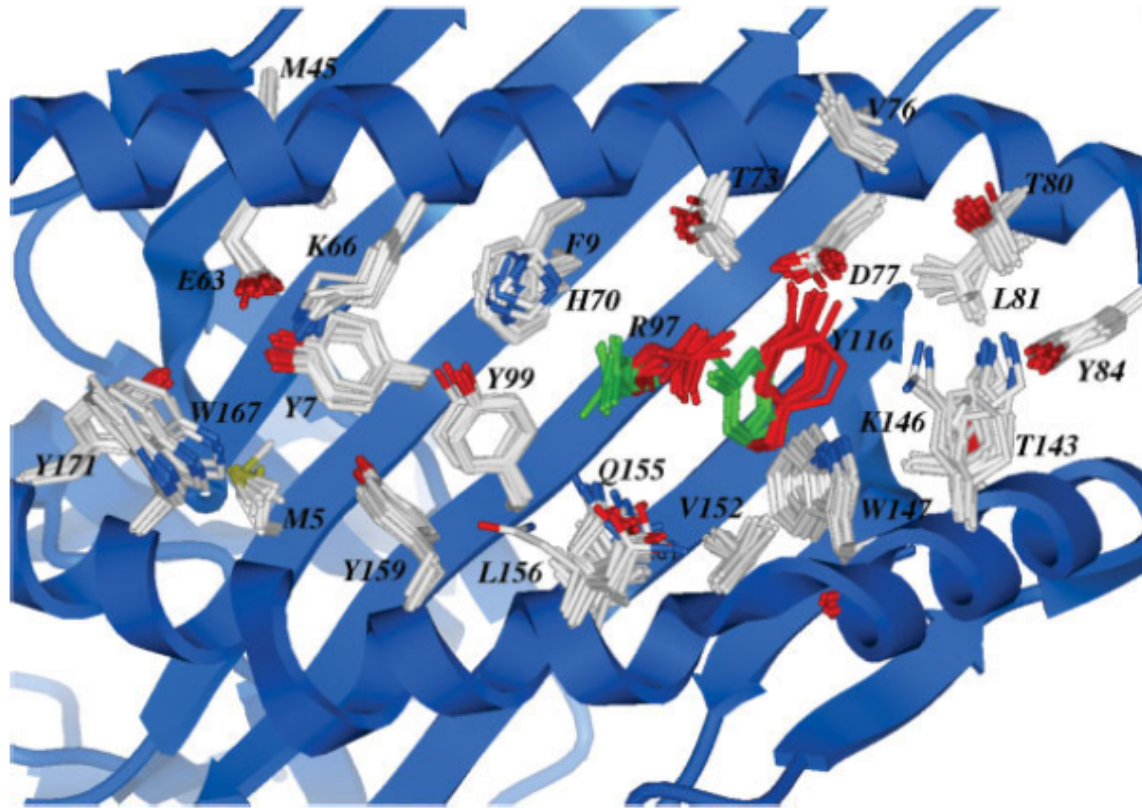
Cross docking of ILSAVGIG



Peptide docking

Ab Initio Prediction of Peptide-MHC Binding Geometry for Diverse Class I MHC Allotypes (4D docking)
 Bordner, Abagyan, *Proteins*, 2006

MHC 1 peptide cross-docking: multiple pocket conformations



HLA-A *0201 peptide binding pocket

- Grids from 2 alternative conformations
- Full BB & SC Sampling until convergence
- N- and C- terminal Hbonds as d.restraints
- Atomic refinement

Bordner AJ, Abagyan R

Ab initio prediction of peptide-MHC binding geometry for diverse class I MHC allotypes.

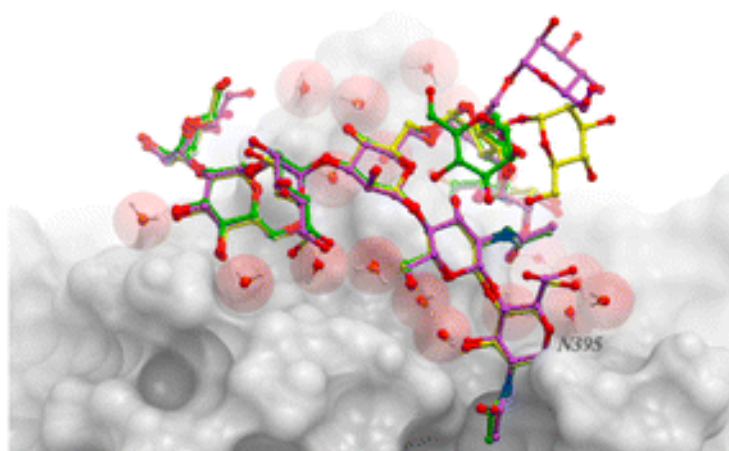
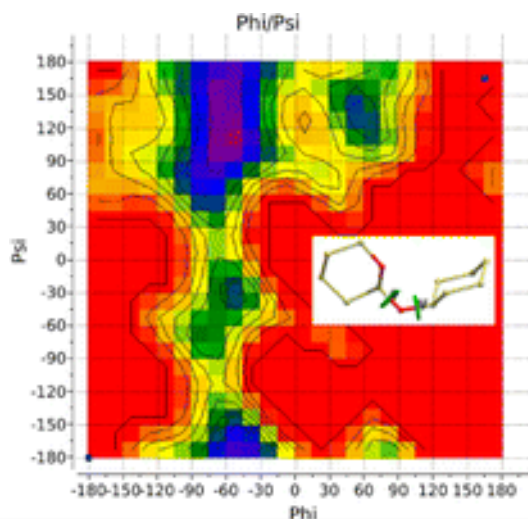
Proteins, 2006 May 15, 63, 512-26

Better ICM Force Field: ICMFF

- First ideas : ICFF *Seva Katrich et al. 2003*
- Current QM-based ICMFF:
 - Softer flexibility model, 3-param VW, $\epsilon=2$, better combination rules, torsion profiles, \therefore

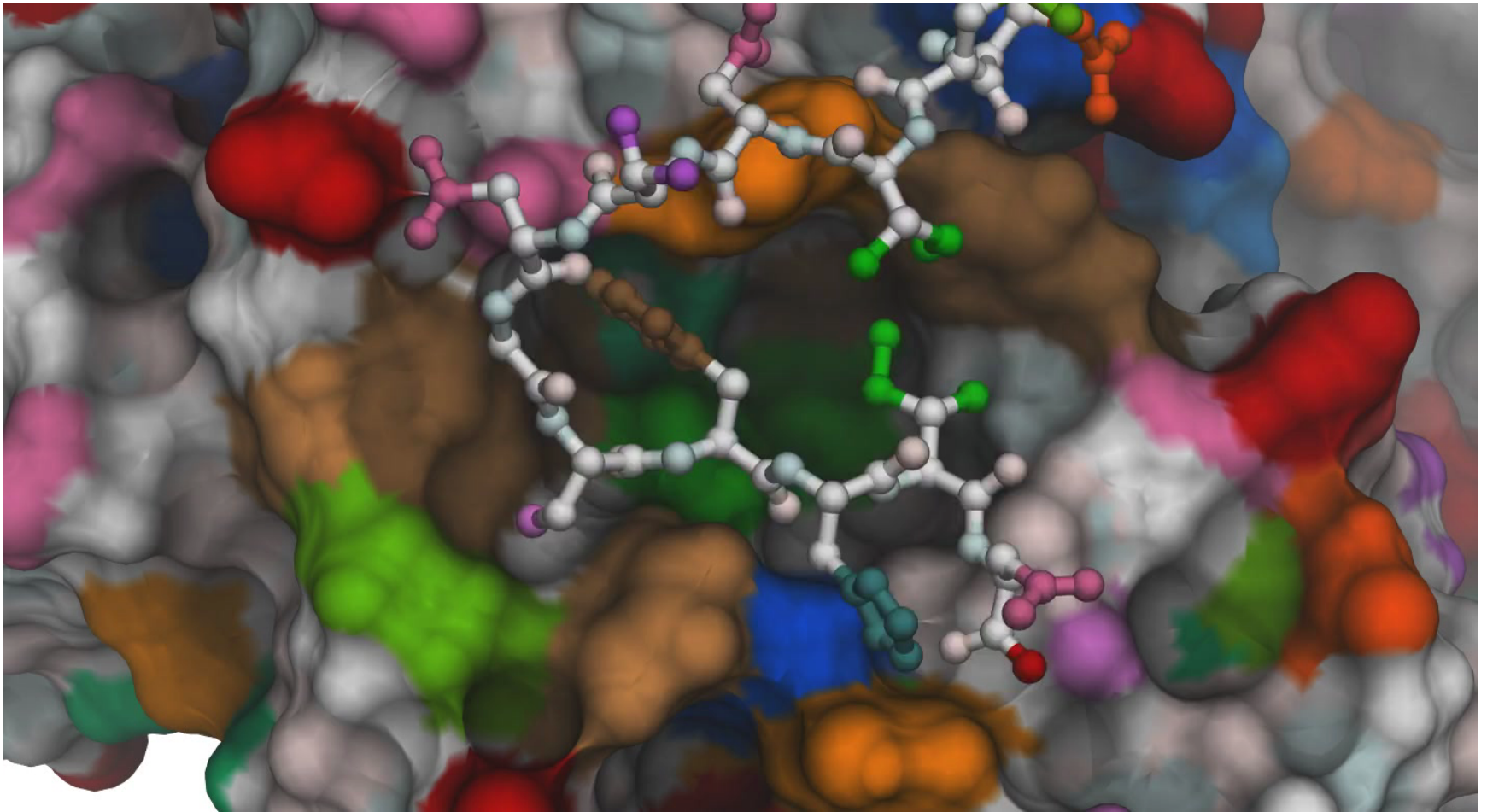
ICM Scripts:

- `_loopmodel`
- `_dockScan`
- `_mutant*`
- `_protDesign`



Arnautova, Abagyan, Totrov. RNA; 2015 (glycoproteins) and 2011 (loops, peptides). **All-Atom Internal Coordinate Mechanics(ICM) Force Field for Hexopyranoses and Glycoproteins.** *J Chem Theory Comput.* 2015

Difficult: 12-residue peptide docking



Performance: 1.5 hours on 1 CPU

Peptide docking benchmark performance

#correct out of 53

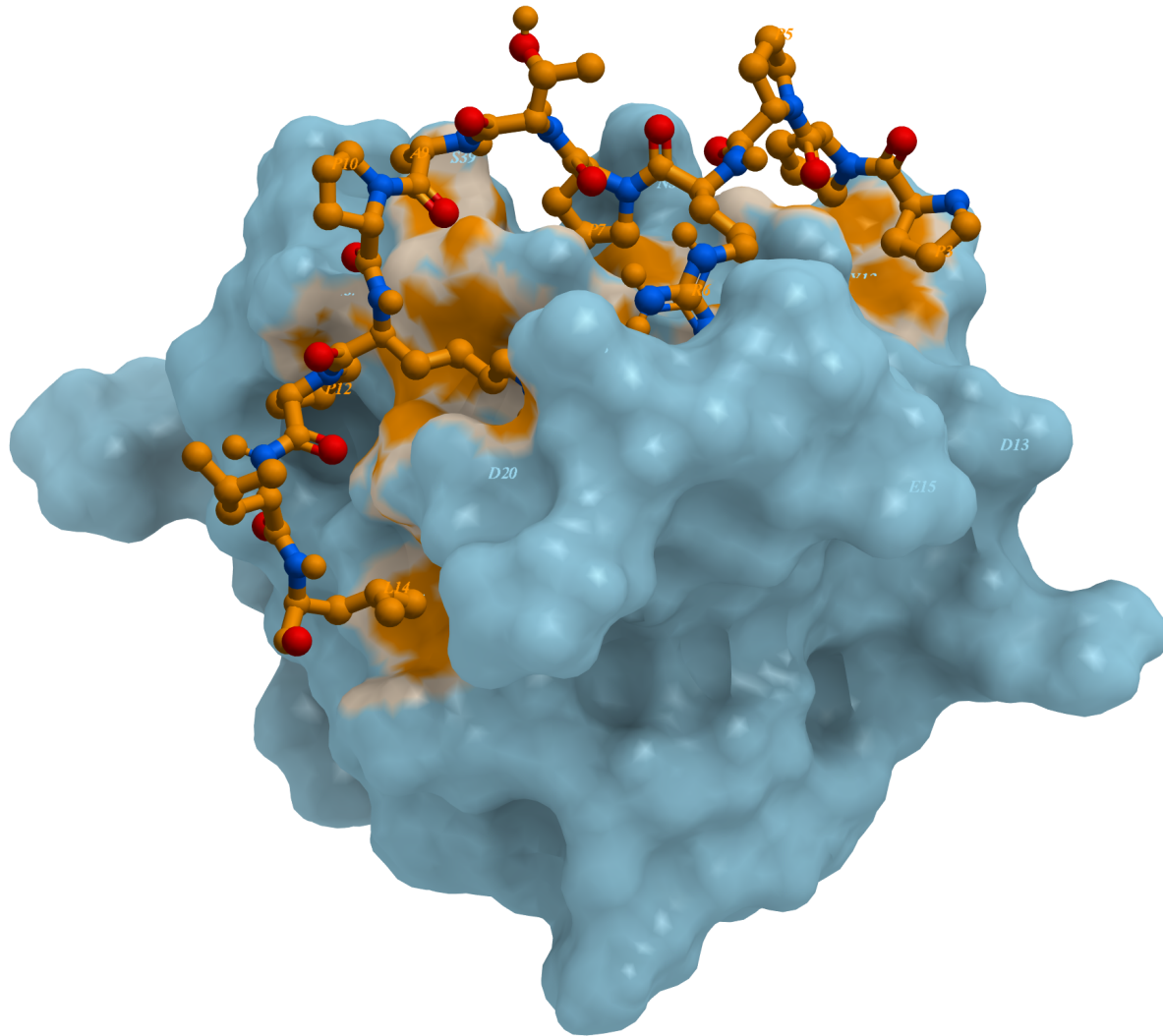
- **AutoDock: 12 /53**
- **Vina: 28 /53**
- **Surflex: 29 /53**
- **GOLD: 28 /53**
- **ICM: 42 /53**

LEADS-PEP: A Benchmark Data Set for Assessment of Peptide Docking Performance
Alexander S Hauser and Björn Windshügel
J. Chem. Inf. Model., 2016, 56 (1), pp 188–200

Ilatovskiy, Abagyan, 2017 (in preparation)

	pdb	ad1	ad2	vina1	vina2	sur1	sur2	g1	g2	g3	g4	icm1	icm30
	3 1b9j	0.8	0.5	0.9	0.4	0.2	0.3	0.3	0.2	0.3	0.4	0.3	0.2
2	3 2oy2	0.5	0.4	1.1	0.8	1.3	5.5	0.5	0.5	0.4	0.4	1.2	1.2
3	3 3gq1	0.9	1.4	1.3	0.9	0.5	0.3	0.8	0.4	0.6	0.6	0.8	0.8
4	3 3bs4	3.3	0.5	0.6	0.4	0.4	0.4	0.7	0.3	0.4	0.4	0.3	0.3
5	3 2oxv	1.3	0.8	1.1	1.1	2.5	5.3	0.7	1.0	0.7	2.2	1.6	1.6
6	3 2b6n	7.6	7.7	0.5	5.2	7.5	7.8	2.3	2.7	2.2	1.3	4.4	1.7
7	4 1tw6	1.0	0.8	0.7	1.0	0.7	0.7	0.6	0.4	0.4	0.4	0.6	0.3
8	4 3vqg	2.6	2.8	0.3	0.6	0.4	0.3	0.6	5.8	0.8	0.5	0.8	1.0
9	4 1uop	1.1	0.6	2.9	0.4	0.4	2.6	0.7	3.7	4.3	0.4	0.9	0.7
10	4 4c2c	0.9	0.8	0.5	0.6	0.5	0.6	0.6	0.8	0.5	0.4	0.8	0.8
11	4 4j44	0.9	0.7	0.4	0.3	0.8	0.7	0.3	0.4	0.3	0.4	1.2	1.2
12	5 2hpl	1.3	2.3	2.1	1.9	1.1	0.8	2.8	4.5	2.5	4.8	0.9	0.8
13	5 2v3s	3.6	3.7	1.0	0.9	7.5	7.7	1.4	1.3	1.5	0.7	3.0	0.9
14	5 3nfk	2.8	3.3	4.8	1.2	0.3	0.3	1.0	2.7	3.3	1.8	0.5	0.4
15	5 1nvr	4.1	3.9	2.7	1.1	2.7	2.4	4.5	6.2	6.2	4.2	1.0	1.0
16	5 4v3i	4.8	5.0	5.2	5.2	3.4	2.8	2.0	1.6	1.2	2.0	5.3	5.2
17	5 3t6r	4.8	2.4	4.0	4.5	4.1	2.7	2.1	0.6	1.7	0.7	0.8	0.5
18	6 1svz	3.4	5.1	3.7	0.7	2.3	2.3	6.2	3.5	4.3	2.9	1.2	1.2
19	6 3d1e	1.3	0.4	3.3	4.0	1.7	1.8	4.6	1.4	0.4	3.1	1.9	1.5
20	6 3idg	4.9	4.5	3.5	2.3	1.2	5.0	4.8	5.8	3.8	4.9	3.0	2.7
21	6 3lny	5.0	6.2	0.6	0.9	6.7	11.0	0.6	0.7	0.6	1.2	0.7	0.8
22	6 4nnr	5.7	5.4	0.9	0.8	0.7	0.9	2.3	1.5	0.9	1.4	2.8	1.1
23	6 4q6h	5.1	6.0	7.8	6.8	6.3	1.3	3.4	2.9	1.6	2.6	0.4	0.6
24	7 3mm	7.6	8.2	5.7	1.2	0.7	0.5	7.7	4.8	2.9	1.3	0.8	0.9
25	7 3q47	4.3	6.0	3.3	1.6	6.1	3.2	5.4	4.5	0.7	1.4	1.8	1.0
26	7 3upv	3.0	4.7	1.5	0.6	1.6	2.2	3.8	3.5	5.1	3.4	3.2	0.4
27	7 4qbr	7.9	4.5	2.1	1.2	0.6	1.0	5.5	3.2	8.6	1.8	1.1	0.9
28	7 3njg	9.2	2.5	1.2	1.3	0.9	0.4	3.2	2.6	1.2	1.1	0.6	0.3
29	8 1elw	5.2	3.2	3.8	2.6	2.0	2.5	3.5	3.4	3.0	1.7	2.1	2.6
30	8 3ch8	7.9	6.3	5.2	0.5	1.5	4.2	6.3	8.5	7.7	3.9	3.3	0.4
31	8 4wlb	6.0	5.3	5.2	5.2	5.1	4.7	3.5	5.2	3.7	4.0	3.6	4.1
32	8 1ou8	6.0	4.1	5.3	4.5	1.3	1.4	3.6	3.6	4.9	3.6	2.0	0.6
33	8 1n7f	4.6	6.3	8.5	11.7	9.1	8.9	6.0	7.5	6.1	8.1	0.9	1.0
34	9 3obc	5.9	10.0	7.4	5.5	1.8	2.1	2.9	2.2	3.0	1.4	2.2	1.8
35	9 4btb	1.9	4.7	2.4	2.2	8.6	8.5	4.3	6.8	3.4	5.9	1.8	1.6
36	9 2w0z	6.3	6.8	5.1	3.8	1.3	4.1	3.2	1.4	3.0	3.3	11.1	1.9
37	9 4n7h	3.8	5.0	4.8	4.7	3.4	4.5	3.8	5.3	6.8	2.2	3.5	1.7
38	9 2qab	3.8	4.3	4.0	4.0	4.6	4.5	3.6	4.1	3.7	4.6	3.8	4.3
39	10 1h6v	11.9	13.1	5.6	3.2	0.6	1.3	13.8	16.4	13.2	1.5	1.6	1.5
40	10 3brl	8.1	7.8	6.0	4.4	3.2	2.4	3.4	4.0	3.6	2.5	3.8	0.8
41	10 1ntv	5.9	3.8	5.4	3.7	15.1	13.4	2.9	5.7	4.1	4.3	1.0	0.4
42	10 4ds1	6.7	5.5	5.9	2.8	2.2	1.6	3.5	4.6	2.8	2.3	1.2	0.5
43	10 2o02	3.7	5.0	4.3	4.2	4.6	3.8	4.6	3.6	4.2	3.7	1.4	0.7
44	11 1n12	11.9	11.4	10.5	9.6	0.5	1.3	8.9	11.7	9.3	2.3	1.6	0.5
45	11 2xfx	7.4	8.0	4.7	2.0	6.4	1.3	4.3	6.1	4.7	4.5	1.7	0.9
46	11 3bfw	7.9	10.0	6.1	5.8	0.3	0.3	6.0	5.8	4.8	3.5	0.3	0.3
47	11 4eik	7.9	5.3	2.2	1.5	4.0	2.5	3.6	3.2	3.9	3.5	3.7	0.9
48	11 3ds1	5.0	5.0	4.5	4.5	11.5	10.6	4.2	6.2	5.4	6.7	4.1	1.2
49	12 4j8s	4.9	5.1	4.7	8.4	5.7	13.2	4.4	5.5	4.1	8.7	9.2	6.7
50	12 2w10	8.5	5.8	2.1	4.6	3.5	3.7	7.3	2.3	3.5	5.6	4.0	2.7
51	12 3jzo	5.7	5.8	5.2	5.1	5.8	5.9	6.4	4.9	5.1	5.2	6.5	7.4
52	12 4dgy	5.7	5.6	7.4	1.1	7.7	7.7	8.9	7.9	5.0	5.8	2.8	5.6
53	12 2b9h	6.1	5.2	6.0	5.5	12.7	10.0	3.4	4.2	3.8	3.8	3.4	4.0

Peptide Docking example: 12-aminoacid peptide 2w10



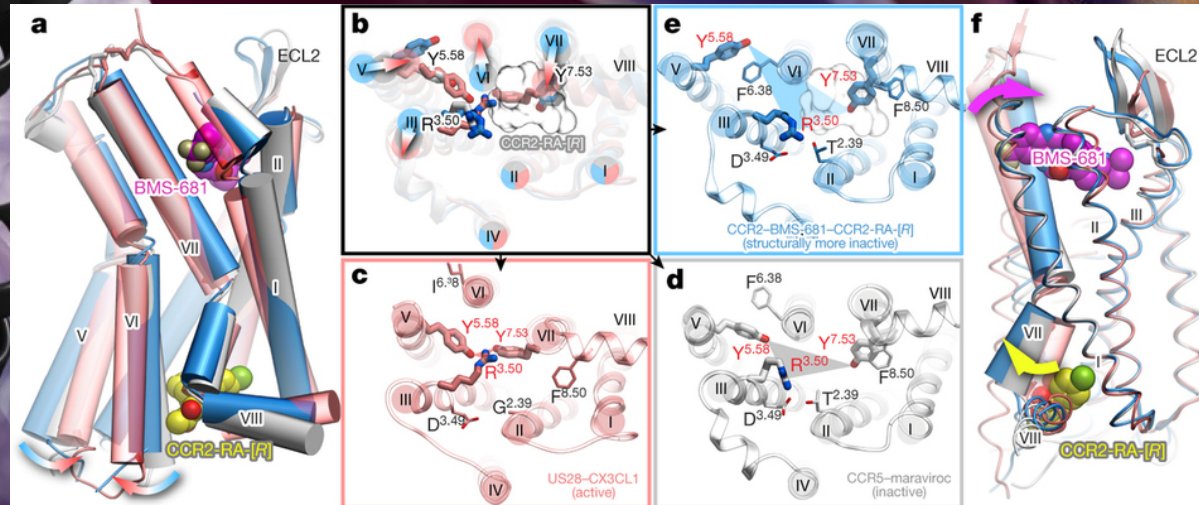
Chemokine

CXCR4

Qin L, Kufareva I, Holden LG, Wang C, Zheng Y, Zhao C, Fenalti G, Wu H, Han GW, Cherezov V, Abagyan R, Stevens RC, Handel TM
Structural biology. Crystal structure of the chemokine receptor CXCR4 in complex with a viral chemokine.
Science, 2015

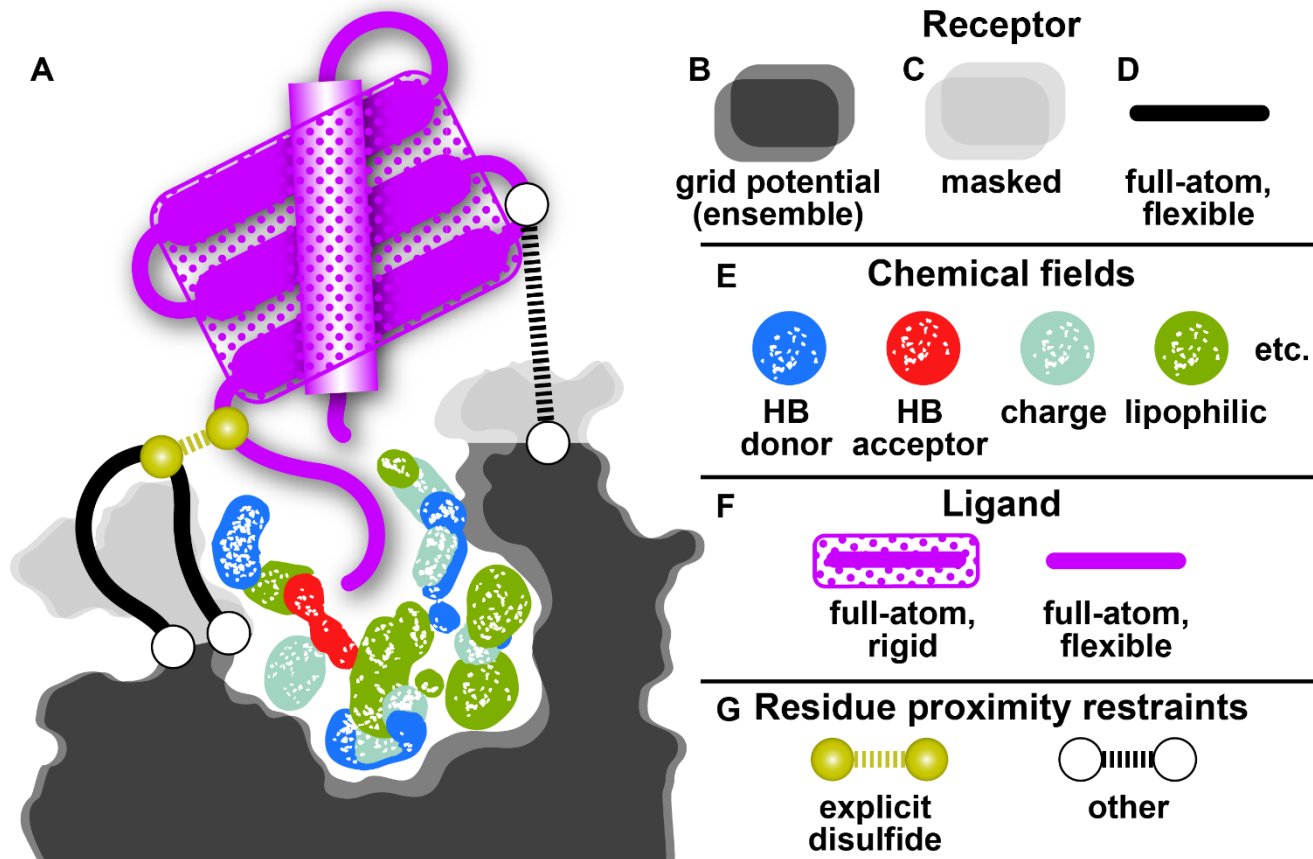
Structure of CC chemokine receptor 2 with orthosteric and allosteric antagonists. Zheng et al.
Nature 2016

Structural basis of ligand interactions with atypical chemokine receptor 3.
Gustavsson et al. *Nat. Commun.* 2017

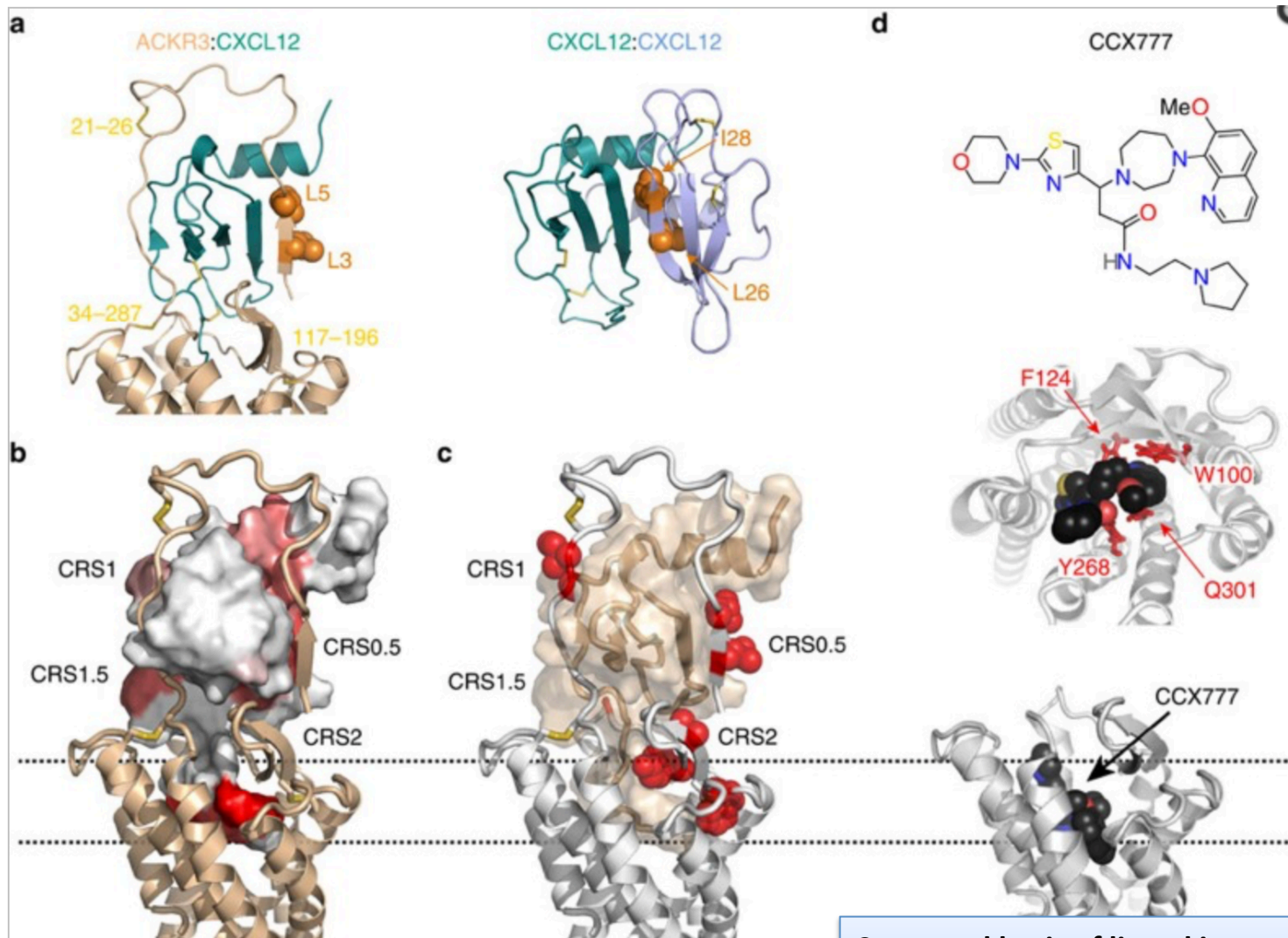


Hybrid modeling & docking protocols

- Stochastic optimization of a system in internal coordinates (ICM)
- Explicit Flexibility of tails, loops and side chains, plus Masking
- Integrated *ambiguous* experimental restraints

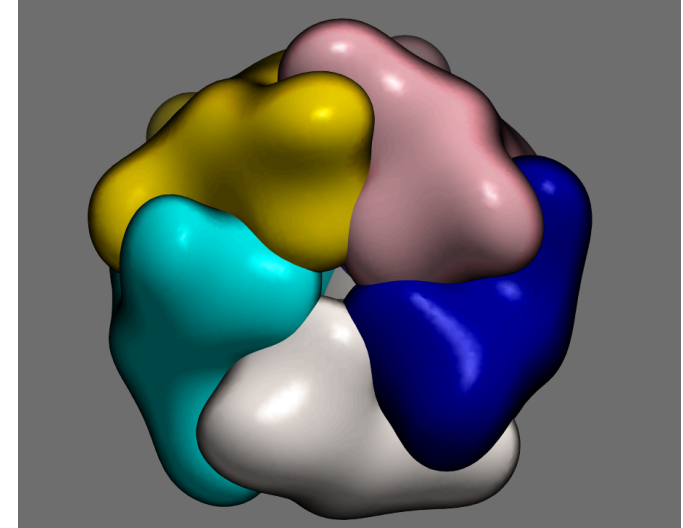
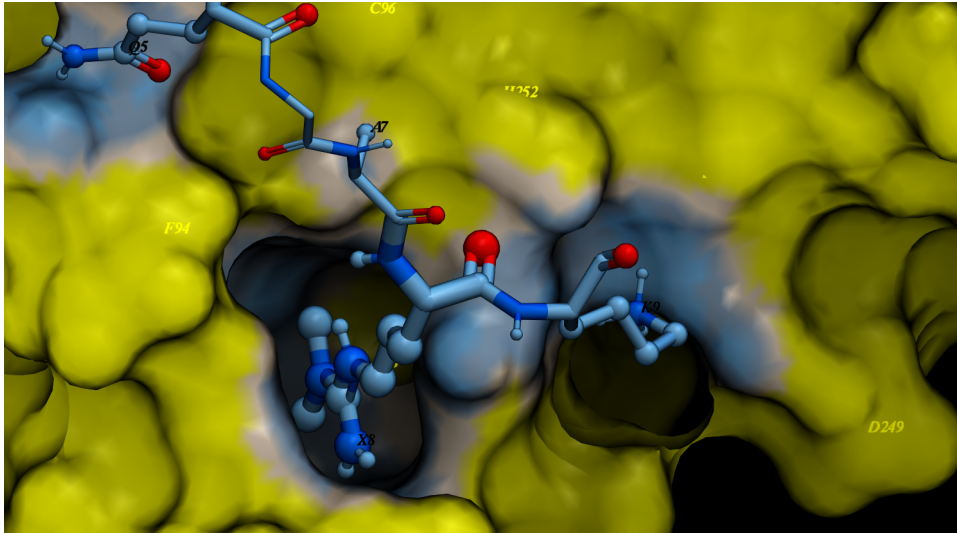


ACKR3-Nterm and CxCL12



Structural basis of ligand interaction with atypical chemokine receptor 3. Gustavsson et al.,
Nature Comm, 2017 (collaboration with Handel Lab)

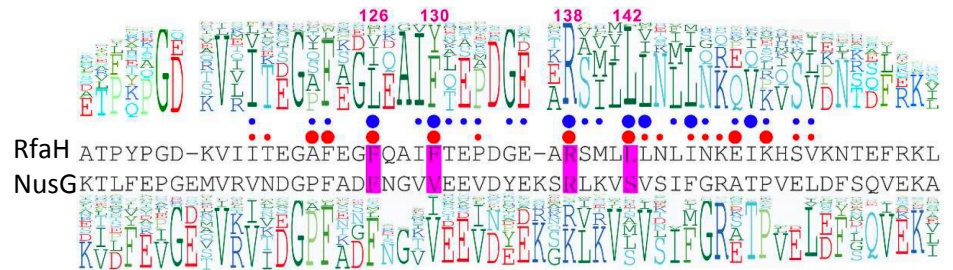
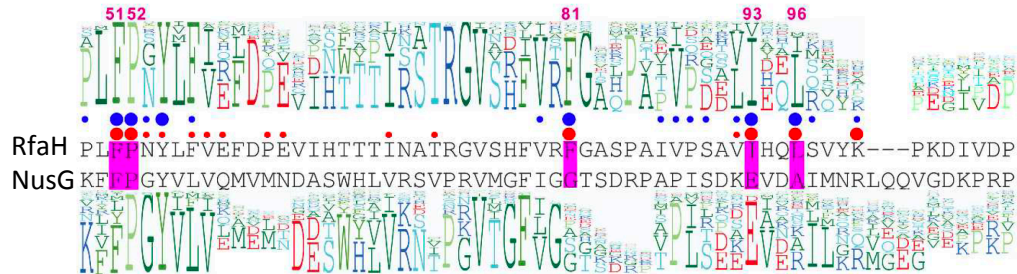
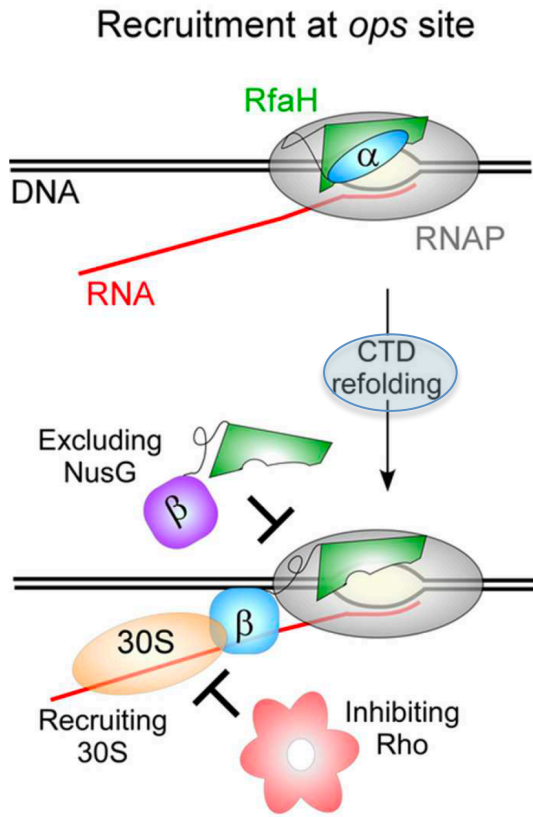
Homology Modeling



- Single mutations (geometry, stability, ppi, ligand binding)
- Search for template(s)
- Structural alignments/edits
- Including Ligand
- Loops & Ends
- Refinement

ID=20%	pP=8.4KLP	PPGPT.#P	IG.....D	#+.#	YG.##	..K.##	L#.Y-#																																																																																																																																										
cyps	1	MWTILLSTIN	ITLATALM	LSFII	IYLYIQ	NSTKL	PPGPT	SWPLIG	YTSCLGT	-DAFR	KIQD	LNKI	YGDIV	SFQV	LKGT	IIII	LYNDL																																																																																																																																	
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Inr6	a	57	V	K	E	A	L	V	D	G	E	E	F	A	G	R	G	S	V	P	I	L	E	K	V	S	K	G	L	G	I	A	-	F	S	N	A	K	T	W	K	E	M	R	R	E	S	L	M	T	L	R	N	F	M	G	M	K	R	S	I	E	D	R	I	O	E	E	A	R	L	V	E	E	L	R	K	T	N																																																																		
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cyps	255	I	Y	K	T	V	R	O	L	I	D	N	N	V	G	E	H	N	S	D	S	L	L	G	O	L	I	N	D	L	K	I	N	L	T	K	N	D	I	S	R	L	S	F	E	F	M	A	G	T	D	T	S	L	T	W	A	C	D	Y	L	A	R	A	P	I	K	N	F	I	M	E	K	V	K	H	O	K	L	D	V	N	N	P	R	F	I	D	C	F	L	I	K	M	E	O	N	N	L	E	F	T	L	E	S	L	V	I	A	V	S	D	L	F	G	A	G	T	E	T	T	T	L	R	Y	S	L	L	L	K	H	E	V	A	A	R	V	O	E	E	I	E	R	V
Inr6	a	214K	E	S	L	K	S	S	D	L	D	M	T	H	R	W	A	S	V	P	L	S	L	P	H	I	V	R	E	S	F	L	K	N	Y	I	P	K	S	S	I	L	T	N	L	Y	A	V	H	N	S	O	I	K	L	I	N	T	E	O	N	S	D	E																																																																																
cyps	327	I	G	R	H	R	S	P	C	M	D	R	S	R	M	P	Y	T	A	V	I	H	E	O	R	F	I	D	L	L	T	N	L	P	H	A	V	T	R	D	V	R	F	R	N	Y	I	P	K	G	T	D	I	L	T	S	L	T	S	L	H	E	K	A	F	N	P	K	V	F	D	P	G	H	L	E	S	G	N																																																																			
Inr6	a	302K	E	S	L	K	S	S	D	L	D	M	T	H	R	W	A	S	V	P	L	S	L	P	H	I	V	R	E	S	F	L	K	N	Y	I	P	K	S	S	I	L	T	N	L	Y	A	V	H	N	S	O	I	K	L	I	N	T	E	O	N	S	D	E																																																																																
cyps	397	I	O	E	S	D	K	P	I	P	F	S	L	G	S	R	C	P	G	A	R	I	A	N	L	L	E	O	I	L	T	A	N	O	E	F	L	Q	N	I	T	O	S	P	F	E	T	I	S	P	G	N	O	E	S	L	T	P	F	G	I	T	R	T	P	H	K	S	M	Y	I	F	V	T	K	L	N	G	N	R	R	T	S	I																																																														
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Bioinformatics & Modeling



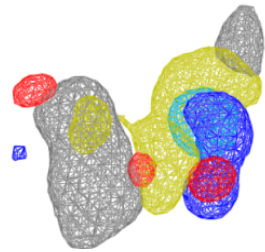
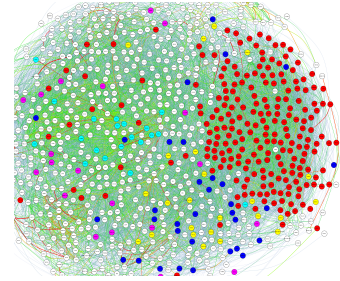
ICM modeling and bioinformatics:

- Alignments of several hundreds sequences for RfaH and NusG
- Calculating Profiles/Logos, Sequence Entropy, Conservation
- Building 3D models of RfaH
- Calculating $\Delta\Delta G$ of mutations for conserved yet different sites
- Prioritizing experimental mutations

Da Shi, Dmitri Svetlov, Ruben Abagyan, and Irina Artsimovitch
Flipping states: a few key residues decide the winning conformation of the only universally conserved transcription factor
Nucleic Acid Research, 2017 (in press)

Conclusions

- Small drugs have **extensive multi-target-pharmacology**, it must and used in matching. We need to use the known and discover the missing.
- **The Pocketome** (~3000) pocket ensembles and superimposed ligand can be used for **Target Screening** via docking combined with machine learning.
- **Complex modeling challenges** can be guided by Internal Coordinate simulations and fuzzy experimental restraints.
- Recent **progress in predictive peptide docking** and scoring enables complex applications.
- **Drug-patient** data helps with targets discovery



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Andrew Orry



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R01 on modeling & docking (active)

U01 on Chemokine Receptors (completed)

UCSD Collaborations

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Tracy Handel lab, *U01, chemokine receptors*

Active Outside UCSD

GPCR structures and modulators

Nicola Smith, Bob Graham, Melbourne, AU

Ray Stevens lab, Seva Katritch USC, U54

Mark Yeager, UVa

Joel Linden's LJIAI, *Adenosine Receptors*

Pat Sexton, **Art Christopoulos, Katie Leach**

Larry Miller lab, Mayo, Arizona, A,B-GPCRs

Multi-target pharmacology

Charles Cunningham, UNM, Bryan Roth,

UNC, Jonathan Marchant

Structure Based Drug Discovery

K.Y. Wong and PolyU team: FtsZ,

glycosyltransferase, TNFa, b-lactamase,

RfaH and NusG

Dmitry Svetlov, Irina Artsimovich

4D docking and activity cliffs